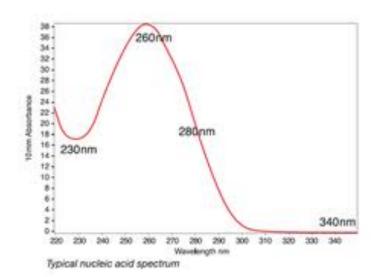
SUN	MON	TUE	WED	THU	FRI	
June 26	27	28	29	30	31	J
	9:00-10:00am Virtual Program Orientation for Summer Institute Online Modality					
July 03	04	05	06	07	08	
	Holiday (Independence Day)	8:30-10:00am -Welcome Reception and Buddy Meet & Greet Event	Free Day	Classes begin! 8:30-11am: BIOL4905 INTRODUCTION 8-10:20pm: Afternoon course	8:30-11am: BIOL4905 DNA PREPARATION 8-10:20pm: Afternoon course	
10) 11	12	13	14	15	
	8:30-11am:BIOL4905 PROTEOMICS I	8:30-11am:BIOL4905 PROTEOMICS II	8:30-11am:BIOL4905 PROTEOMICS III	8:30-11am: BIOL4905 RNA PREPARATION	Virtual Independence	
	8-10:20pm: Afternoon course	8-10:20pm: Afternoon course	8-10:20pm: Afternoon course	8-10:20pm: Afternoon course	Day Activity	
17	18	19	20	21	22	
	8:30-11am:BIOL4905 qPCR / ROBOTS 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 DNA Sequence Analysis 8-10:20pm: Afternoon course	Midterm Break		8:30-11am:BIOL4905 Next Gen. Sequencing 8-10:20pm: Afternoon course	
24	1 25	26	27	28	29	
	8:30-11am:BIOL4905 Microarray I 8-10:20pm: Afternoon course	8:30-11am: BIOL4905 Nanostring 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 Automated Microscopy /AFM	8:30-11am:BIOL4905 Flow Cytometry 8-10:20pm: Afternoon course	FINALS	
3*	August 01	02	03			
	9:00-10:00am: Closing Reception		Grades available in PAWS			





Nucleic acids <u>absorb ultraviolet light in a specific pattern</u>. In a <u>spectrophotometer</u>, a sample is <u>exposed</u> to ultraviolet light at 260 nm, and a photo-detector measures the light that passes through the sample. The more light absorbed by the sample, the higher the nucleic acid concentration in the sample.

The ratio of absorbance at 260 nm and 280 nm is used to assess the purity of DNA and RNA.

A ratio of ~1.8 is generally accepted as "pure" for DNA;

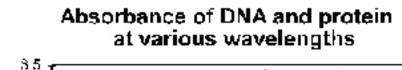
a ratio of ~2.0 is generally accepted as "pure" for RNA.

Unless the there is something strange in the base-composition of the RNA, I would say that your sample is contaminated with a bit of residual Trizol, which is a phenol and guanidium thiocyanate based reagent. I don't know for guanidium thiocyanate, but for sure phenol is known to affect measurements of nucleic acids OD ratios.

L'NA

Protein

DNA/protein



240 245 250 255 290 265 270 275 280 285 200

Wavelength

Absorbance units

Э

2.5

2

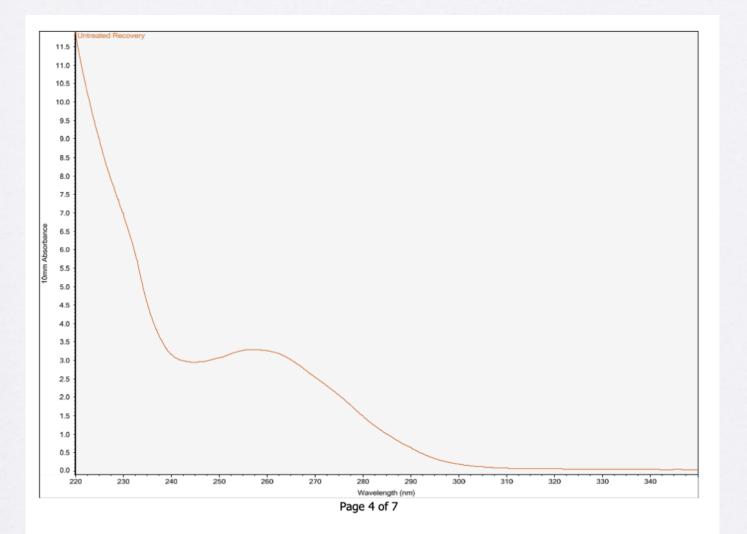
-5

Q.5

υ

115

#	Sample ID	User name	Date and Time	Nucleic Acid Conc.	Unit	A260	A280	260/280	260/230	Sample Type	Factor
1	Untreated 0 Min	Alex Leach	7/20/2022 1:57:52 PM	329.9	ng/µl	8.248	3.691	2.23	1.86	RNA	40.00
2	Untreated 60 Min	Alex Leach	7/20/2022 1:59:17 PM	215.6	ng/µl	5.390	2.411	2.24	1.76	RNA	40.00
3	Untreated Recovery	Alex Leach	7/20/2022 2:00:19 PM	129.0	ng/µl	3.224	1.454	2.22	0.46	RNA	40.00
5	Cadmium 0	Alex Leach	7/20/2022 2:02:11 PM	155.2	ng/µl	3.879	1.750	2.22	0.57	RNA	40.00
6	Cadmium 60	Alex Leach	7/20/2022 2:03:07 PM	492.1	ng/µl	12.301	5.644	2.18	0.72	RNA	40.00
7	Cadmium Recovery	Alex Leach	7/20/2022 2:04:22 PM	782.2	ng/µl	19.556	8.720	2.24	1.91	RNA	40.00



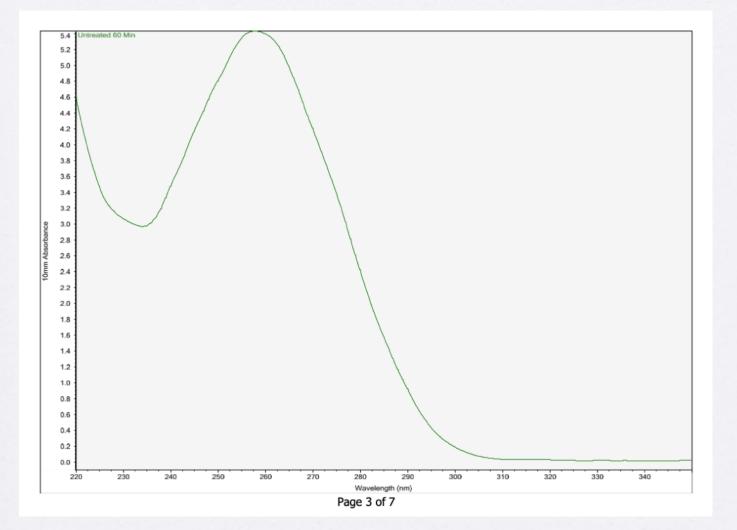
Principle and procedure

RNA purification using RNeasy technology

The RNeasy procedure represents a well-established technology for RNA purification. This technology combines the selective binding properties of a silica-based membrane with the speed of microspin technology. A specialized high-salt buffer system allows up to 100 µg of RNA longer than 200 bases to bind to the RNeasy silica membrane. Biological samples are first lysed and homogenized in the presence of a highly denaturing guanidine-thiocyanate-containing buffer, which immediately inactivates RNases to ensure purification of intact RNA. Ethanol is added to provide appropriate binding conditions, and the sample is then applied to an RNeasy Mini spin column, where the total RNA binds to the membrane and contaminants are efficiently washed away. High-quality RNA is then eluted in 30–100 µl water.

For an OD > 2.0, when purified using the RNeasy technology the RNA sample probably contains higher bits of **residual Trizol**, which is a phenol and **guanidium thiocyanate** based reagent. I don't know for guanidium thiocyanate, but for sure phenol is known to affect measurements of nucleic acids OD ratios.

#	Sample ID	User name	Date and Time	Nucleic Acid Conc.	Unit	A260	A280	260/280	260/230	Sample Type	Factor
1	Untreated 0 Min	Alex Leach	7/20/2022 1:57:52 PM	329.9	ng/µl	8.248	3.691	2.23	1.86	RNA	40.00
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5	Cadmium 0	Alex Leach	7/20/2022 2:02:11 PM	155.2	ng/µl	3.879	1.750	2.22	0.57	RNA	40.00
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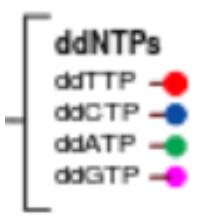
Genes PCR plots ...Low RNA / DNA template concentrations

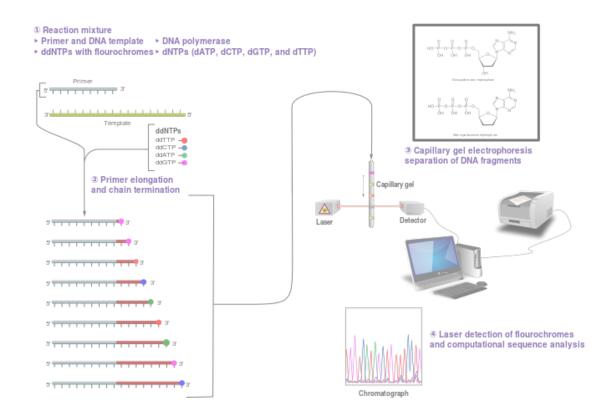


Genetic Analyzer (ABI/ *Life technologies*) Model 3500xl

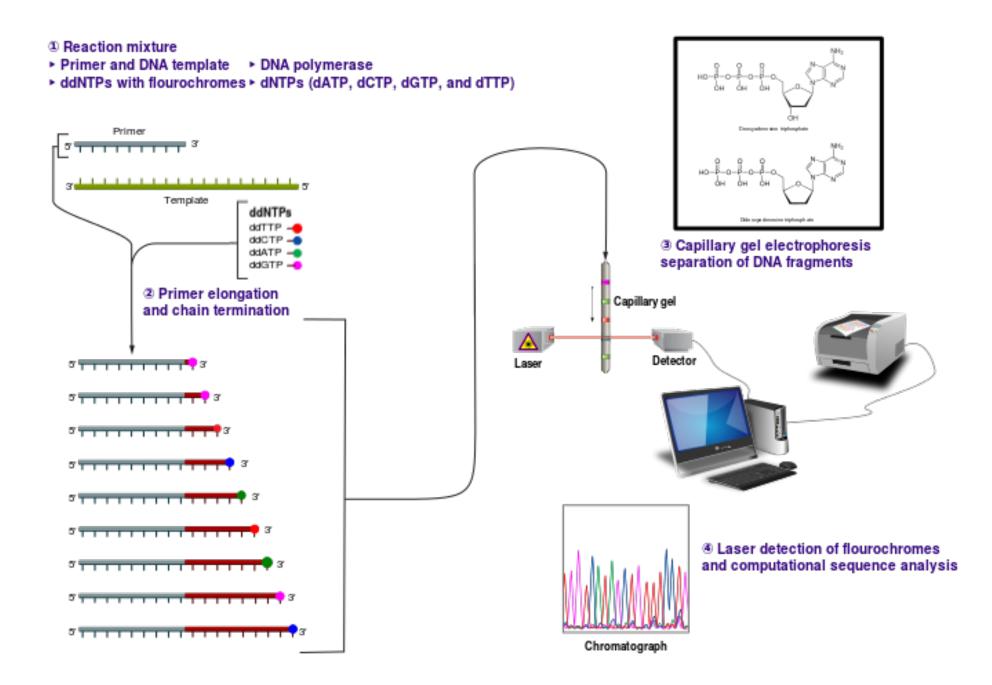
Sanger di-deoxy Sequence Analysis

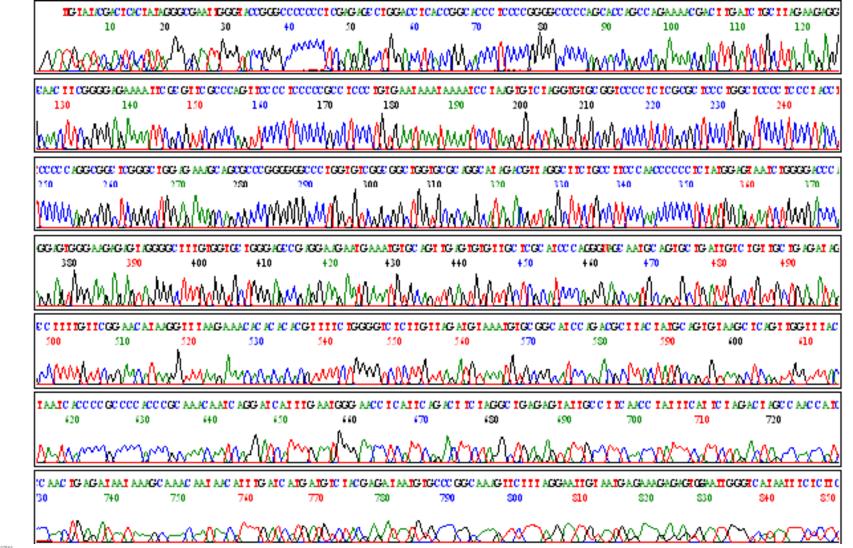






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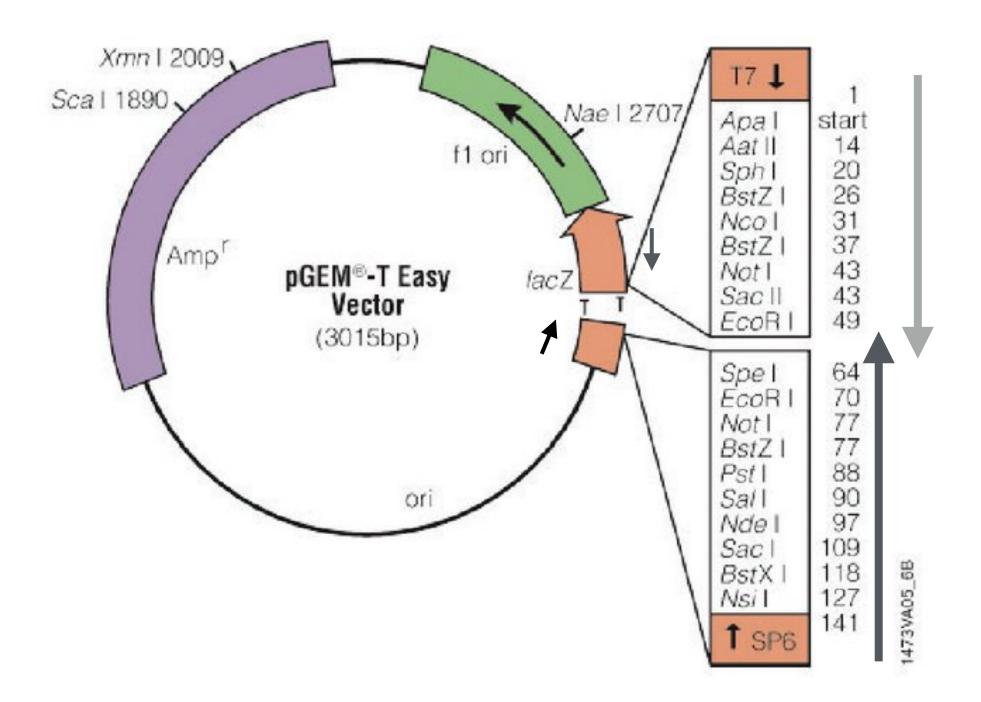


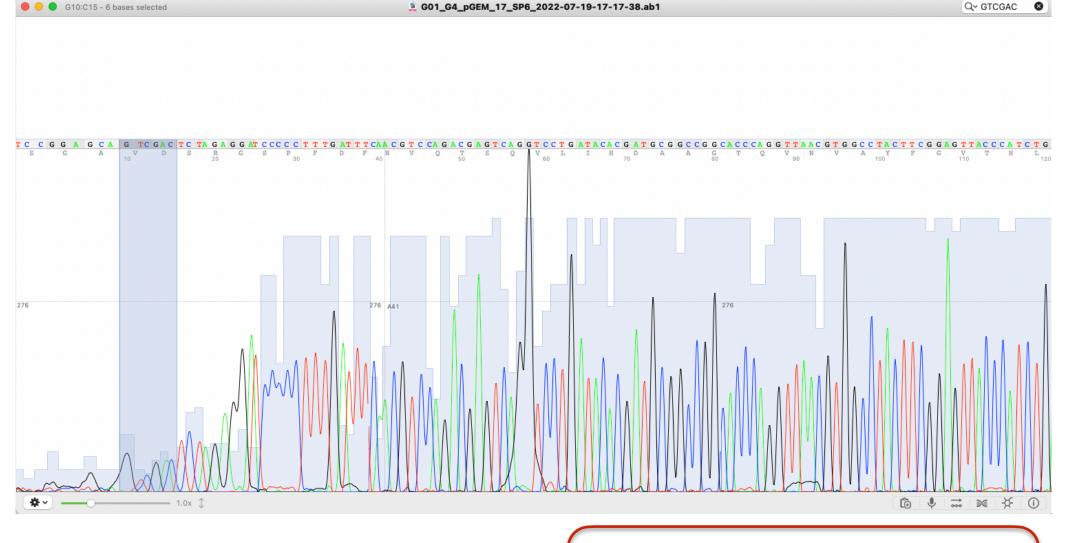
Sanger Sequencing

- Ideal for single gene assays
- Target gene candidates



- · Few amplicons, few samples
- Bidirectional sequencing
- Can be used to confirm variants from PGM





Sall NI 13.1 💥 Ri 🕑 di A 37° 👹 Cog

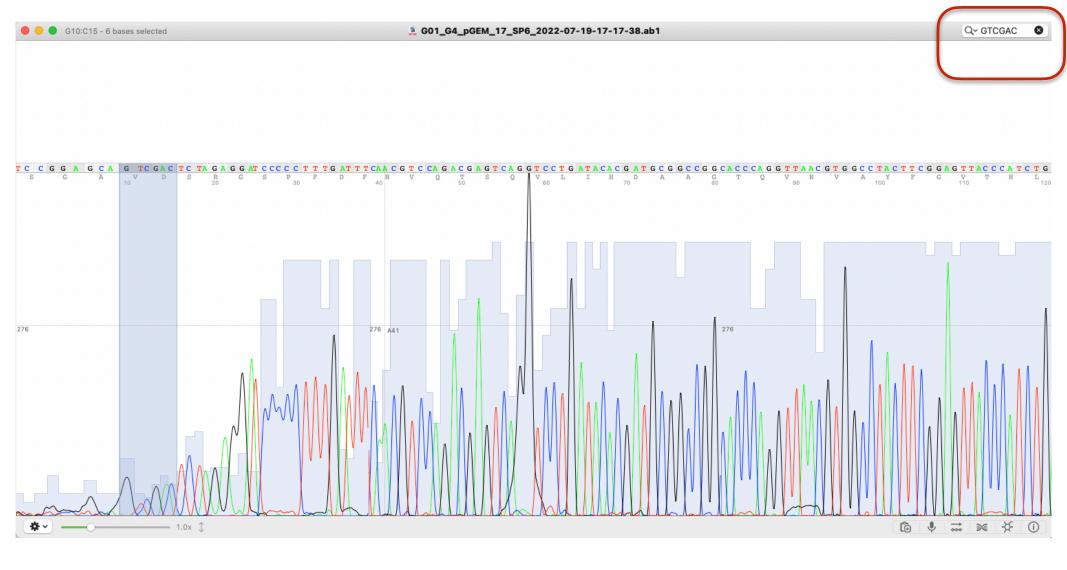
We are excited to announce that all reaction buffers are now BSA-free. NEB began switching our BSA-containing reaction buffers in April 2021 to buffers containing Recombinant Albumin (rAlbumin) for restriction enzymes and some DNA modifying enzymes. Find more details at www.neb.com/BSA-free.

5′... G^TTCGAC...3′ 3′... CAGCTG...5′

Isoschizomers | Single Letter Code | Pronunciation:

- Time-Saver[™] qualified for digestion in 5-15 minutes
- High Fidelity (HF[®]) version available (NEB #R3138) supplied with rCutSmart[™] Buffer
- Supplied with 1 vial of Gel Loading Dye, Purple (6X)
- Restriction Enzyme Cut Site: G/TCGAC

75 Product Citations



Sall 🛛 📽 🕄 🖉 🖬 A 37° 👹 Cog

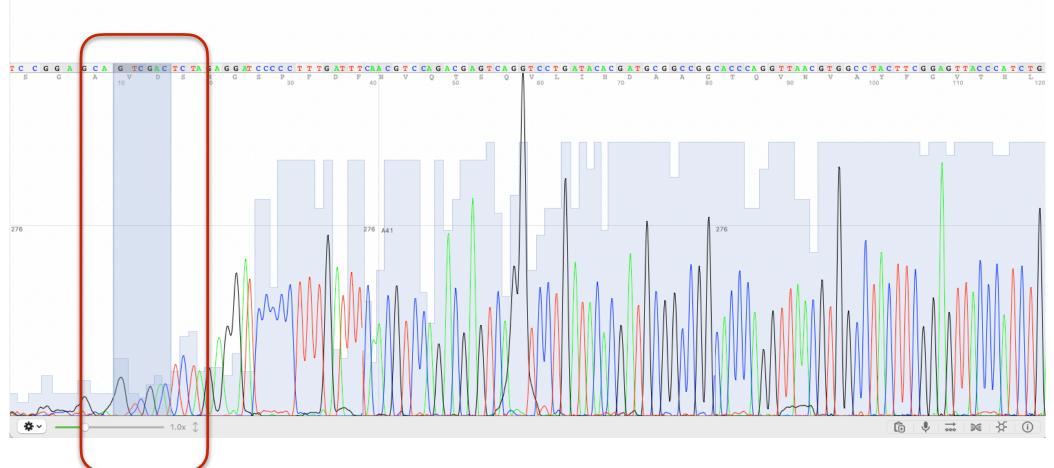
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Sall NE 131 💓 Ri 🖉 di A 37° 👹 CyG

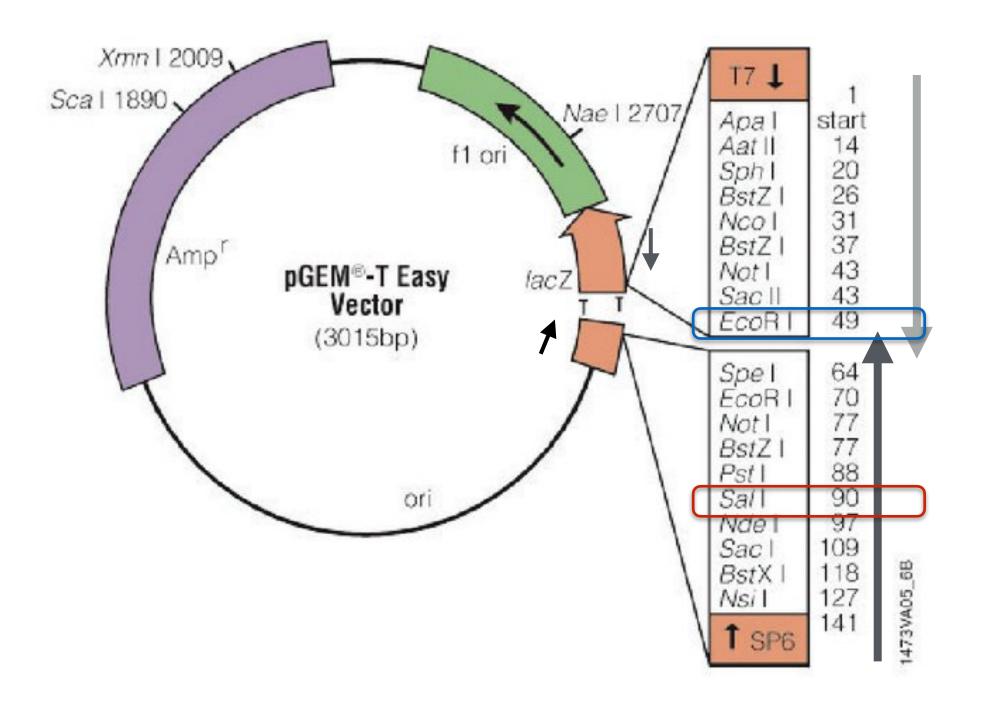
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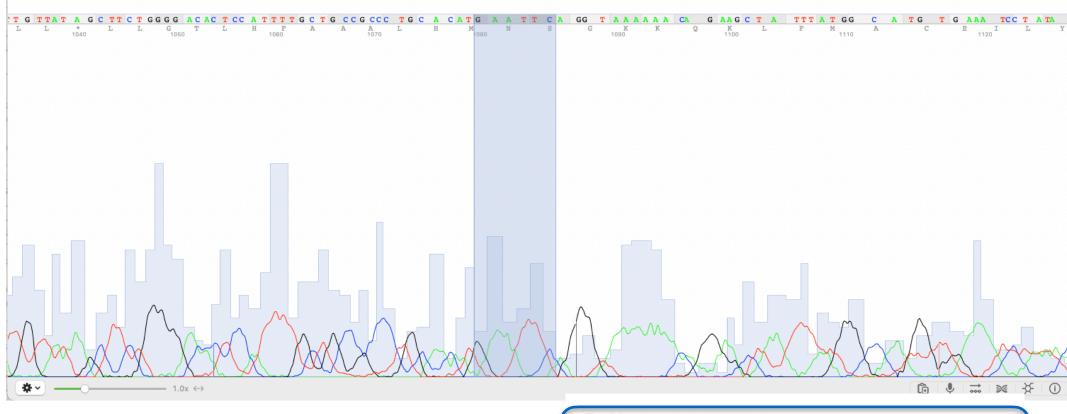
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75 Product Citations





EcoRI KBU 💓 RN 🕐 dic 37° 👹 CpG

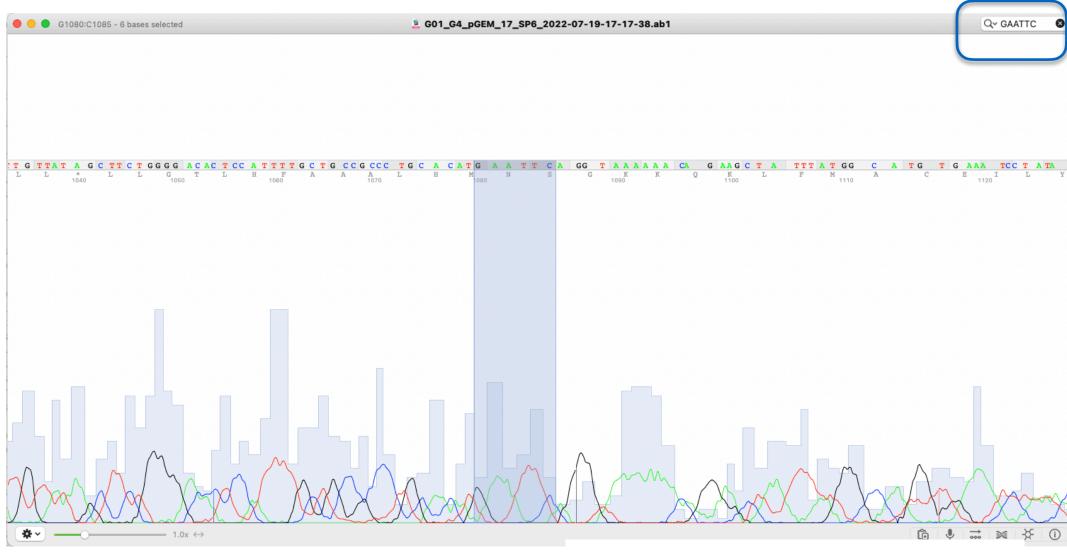
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Isoschizomers | Single Letter Code | Pronunciation:

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- High Fidelity (HF[®]) version available (NEB #R3101) supplied with rCutSmart[™] Buffer
- Now comes supplied with 1 vial of Gel Loading Dye, Purple (6X)
- Restriction Enzyme Cut Site: G/AATTC

2,332 Product Citations



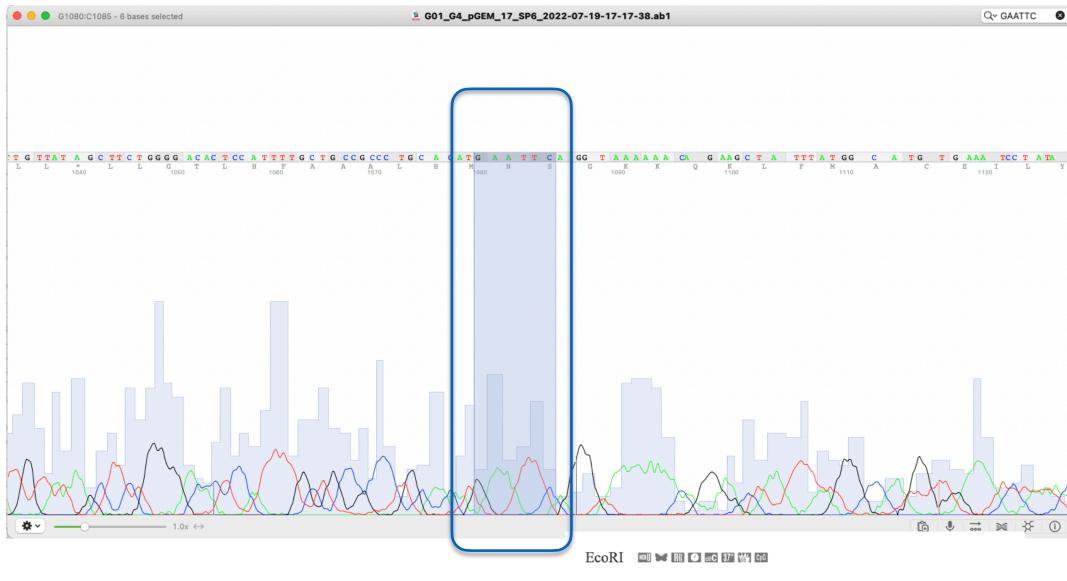
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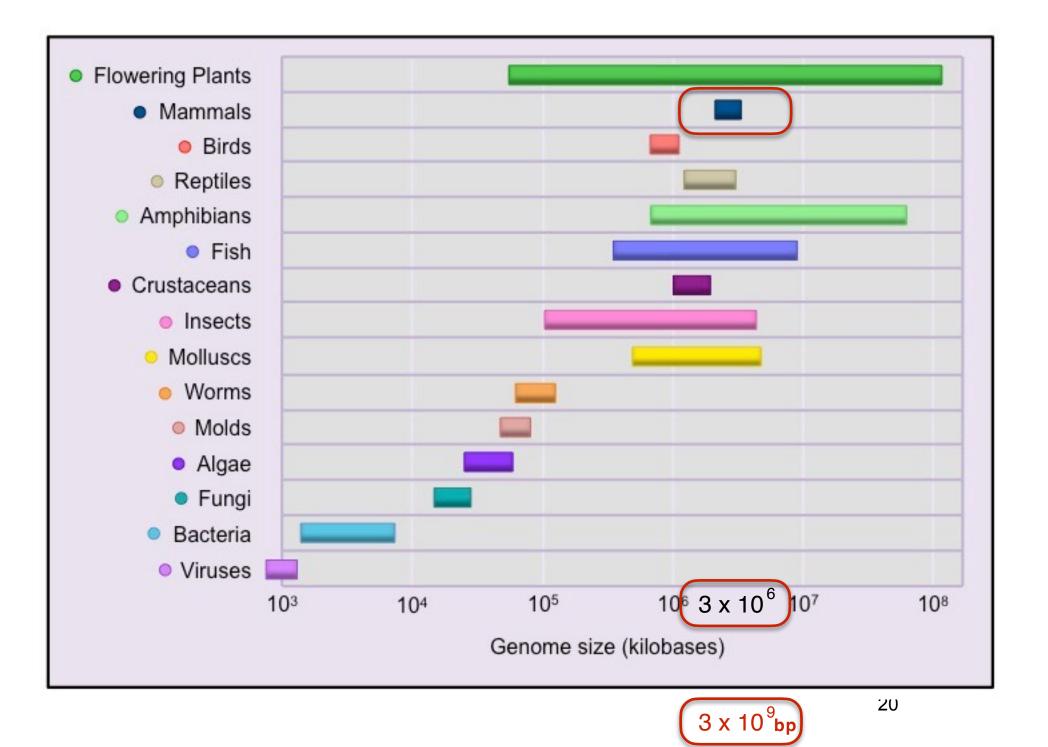
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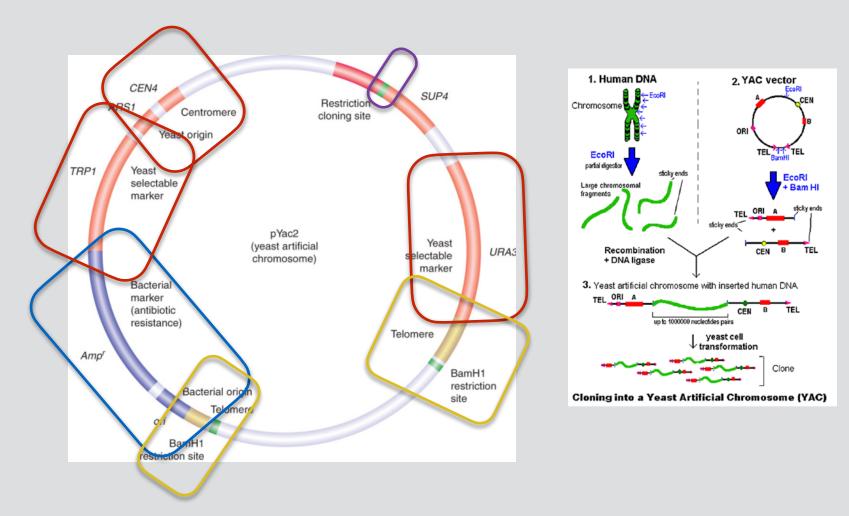
2,332 Product Citations

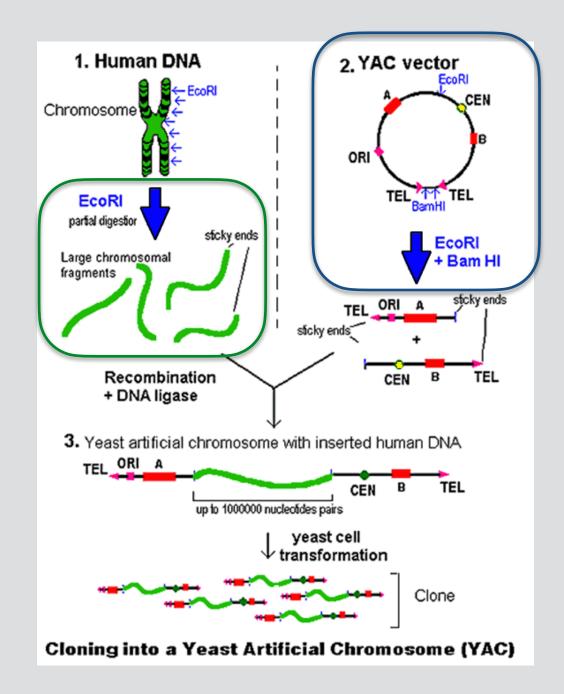


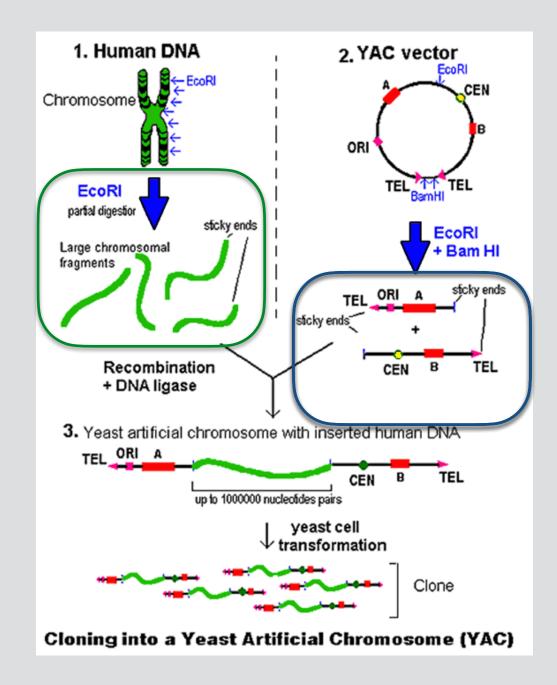
Potential problems:

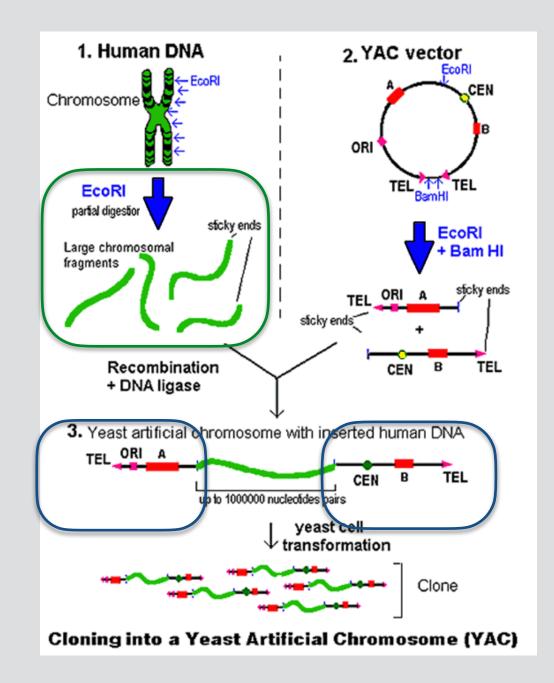
Size of genomes vary among the different 'karyotes', causing logistical problems.

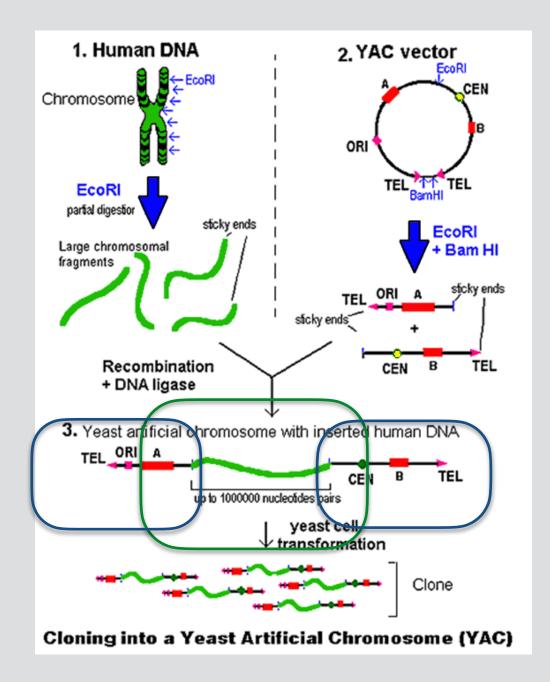
Complexity and Gene structure.

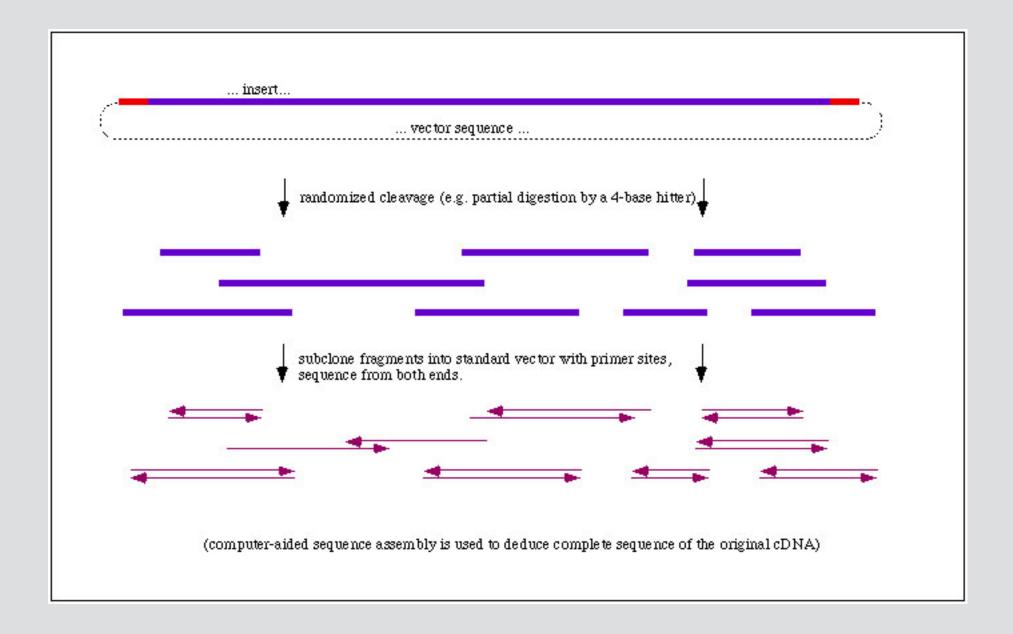














Sanger sequencing throughput

- Run up to 384 samples
- 700 base reads
- Up to 1 million bases per day
- Human genome is 6 billion bases
- Covered 7 times
- Over 100 years on one machine

Next-Generation, Deep Sequencing

Illumina sequencers



	MiSeq	HiSeq	NovaSeq	Sanger
Reads (millions)	30	3,000	13,000	0.0004
Gigabases/day	7	500	4000	0.001
a.gabacco, aay		000	1000	0.001

Next-Generation, Deep Sequencing

Illumina sequencers



	MiSeq	HiSeq	NovaSeq	Sanger
Reads (millions)	30	3,000	13,000	0.0004
Gigabases/day	7	500	4000	0.001
-				

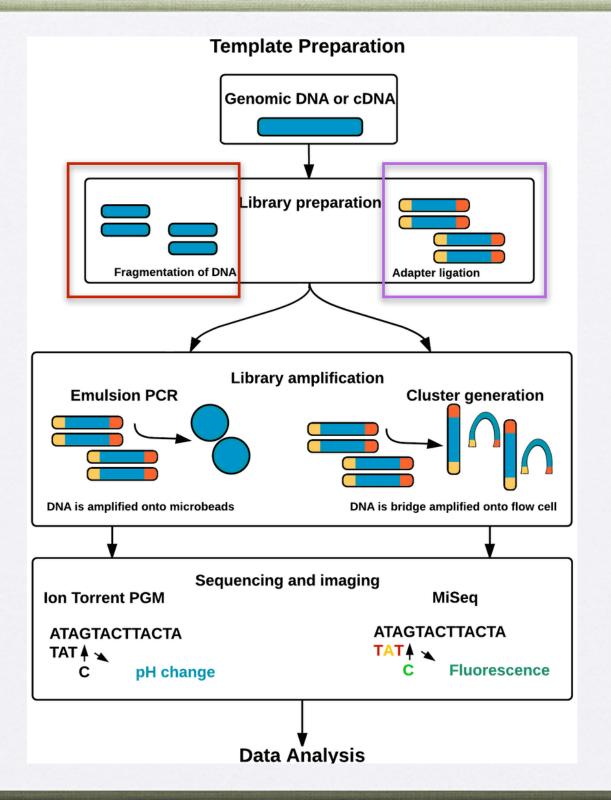
Next-Generation, Deep Sequencing

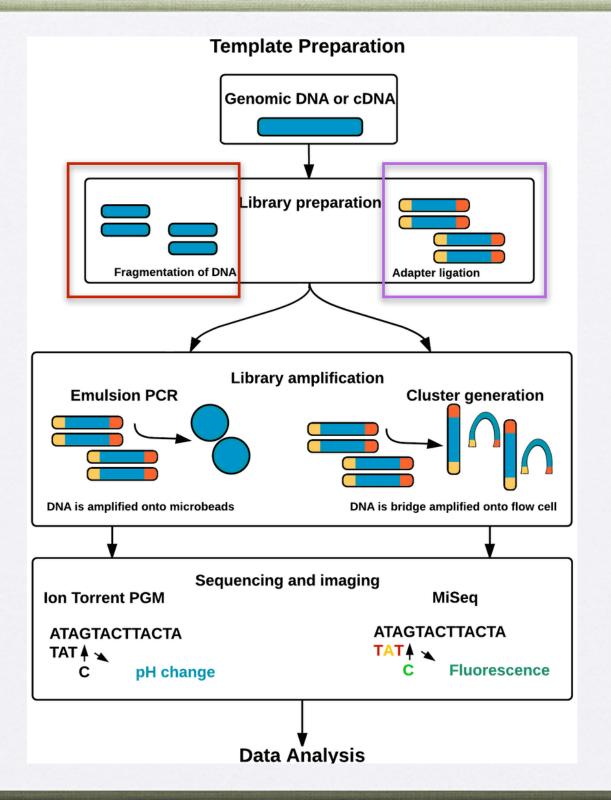


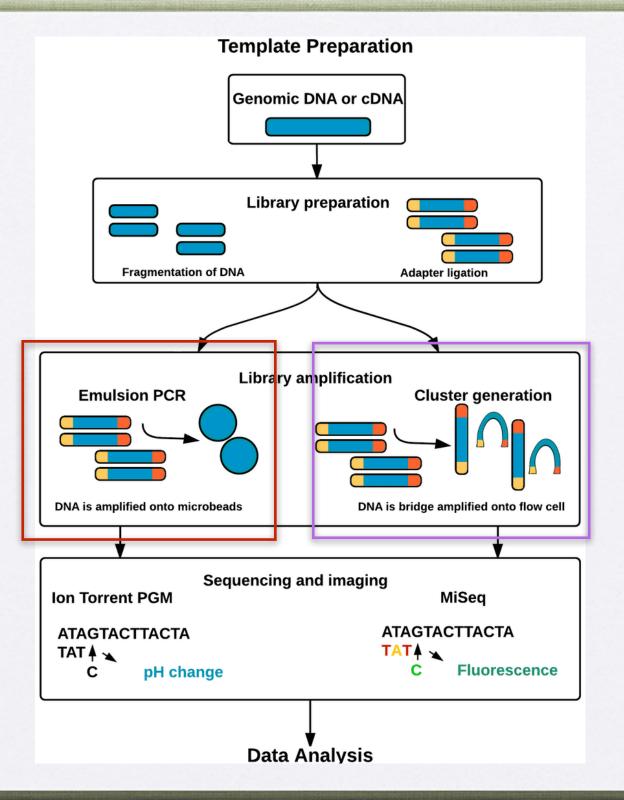
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Gigabases/day	7	500	4000	0.001

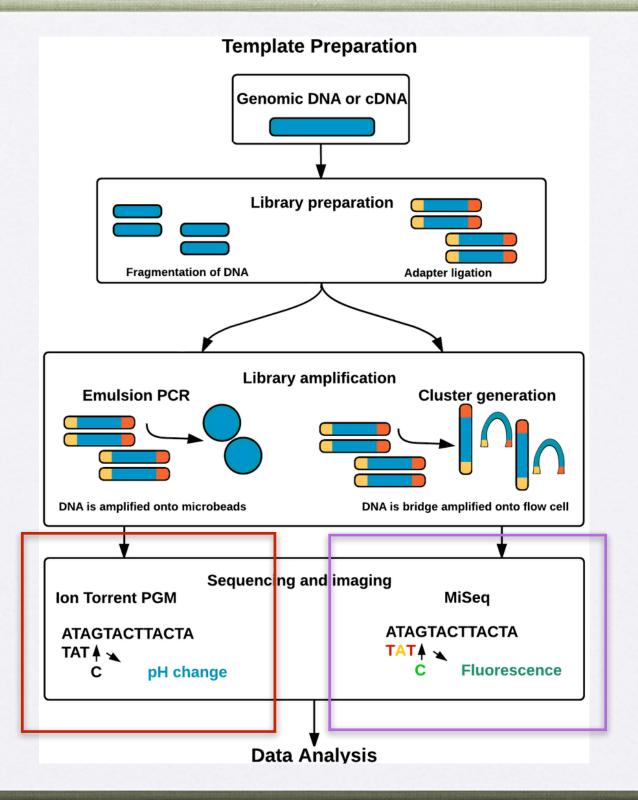


Ion Proton Next-Gen, Deep Sequencing (ABI/Life technologies)









Ion GeneStudio S5 Series I One Platform For All Your RNA Sequencing Needs



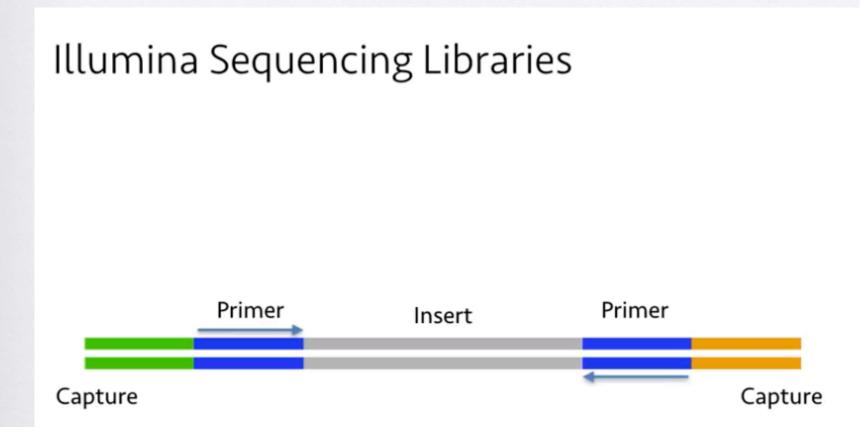
	MiSeq	HiSeq	NovaSeq	Sanger
Reads (millions)	30	3,000	13,000	0.0004
Gigabases/day	7	500	4000	0.001



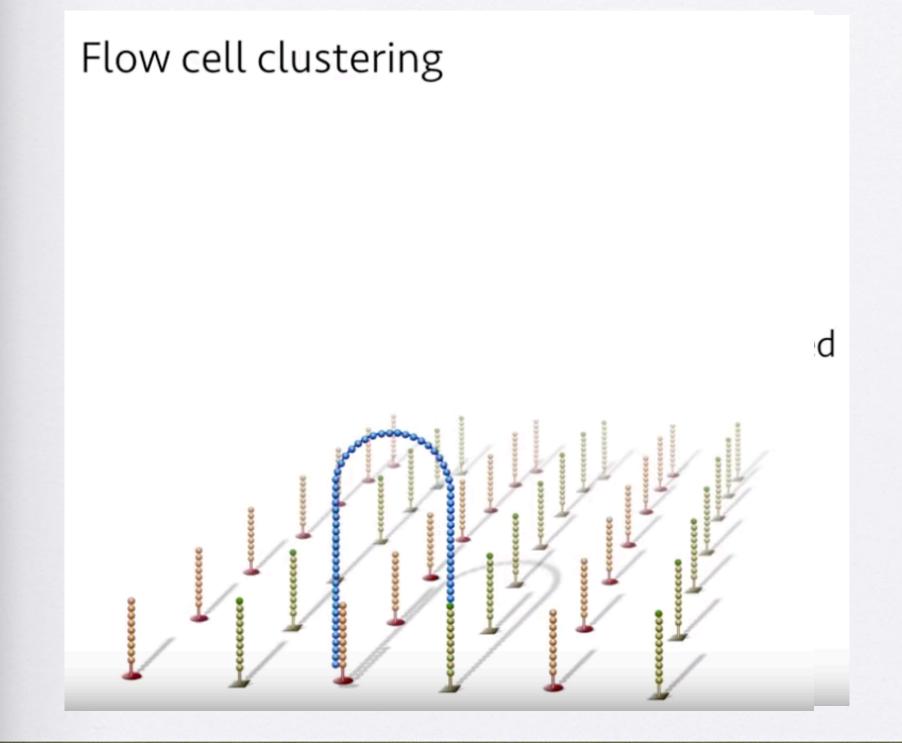
Automation, Ion Chef (ABI/Life technologies)

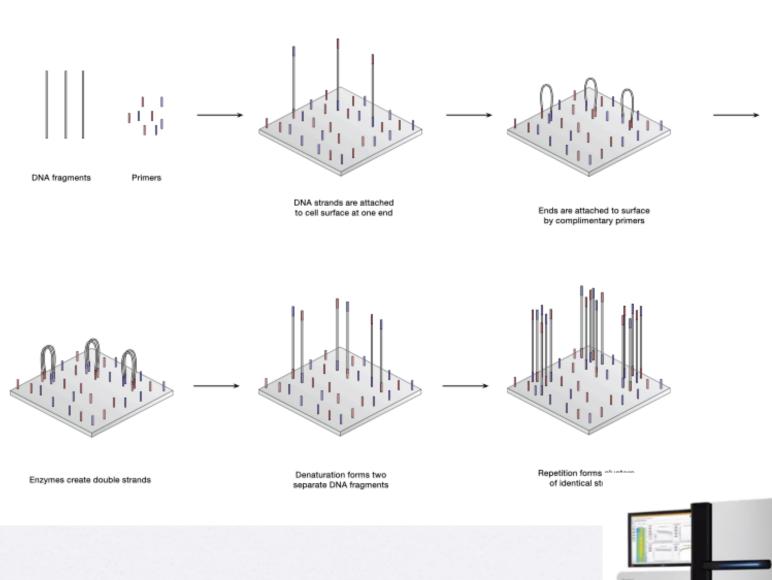


	MiSeq	HiSeq	NovaSeq	Sanger
Reads (millions)	30	3,000	13,000	0.0004
Gigabases/day	7	500	4000	0.001



- Adapters required for sequencing
- Adapter sequence includes primer binding sites and capture sequences.

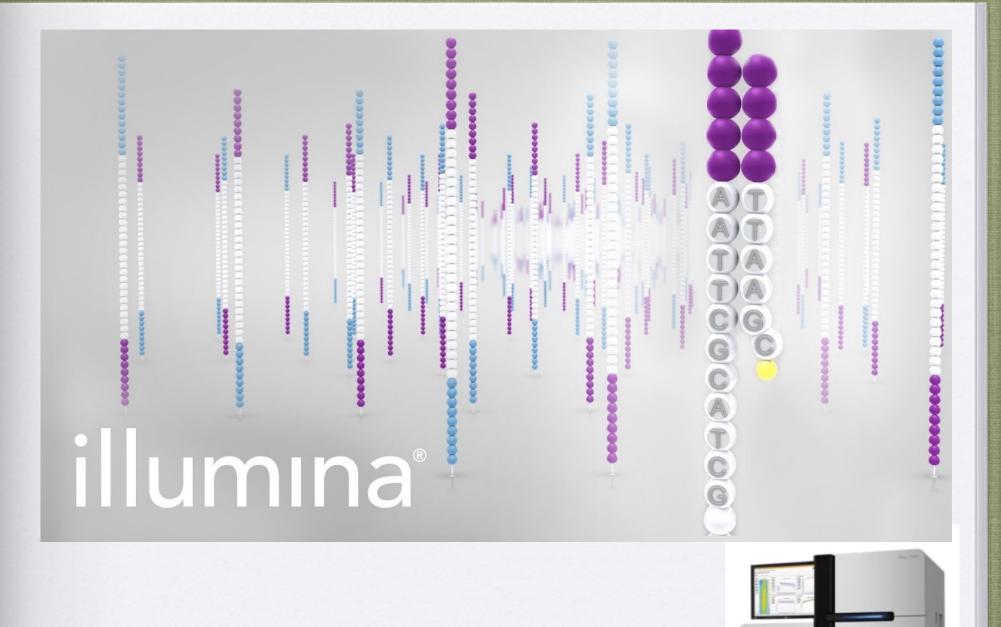




Bridge PCR amplification

Illumina Sequence Analysis





Bridge PCR amplification

Illumina Sequence Analysis

Ion GeneStudio S5 System I Simple. Fast. Scalable. Versatile.

	lon GeneStudio [™] S5	iSeq™	MiniSeq™	MiSeq™	NextSeq™
Scalability ¹	2M, 5M, 20M, 80M, and 130M	4M only	8M and 25M	1M, 4M, 15M, and 25M	130M and 400M
Max throughput per week ²	1,300M reads	20M reads	125M reads	125M reads	1,200M reads
Workflow simplicity ³ (hands-on-time)	45 minutes Automated library prep on Chef	255 minutes Manual library prep			
End-to-end support ⁴	Library prep, Sequencing, Bioinformatics, Reporting	Library prep, Sequencing only	Library prep, Sequencing only	Library prep, Sequencing only	Library prep, Sequencing only
Max read length	Up to 600 bp	Up to 2x150 bp	Up to 2x150 bp	Up to 2x300 bp	Up to 2x150 bp

1. Reads per run

2. Maximum throughput per run multiplied by number of runs per week at highest output: assumes 10 Ion GeneStudio S5 runs of Ion 550 chip per week; 5 runs per week of iSeq[™], MiniSeq[™], and MiSeq[™] sequencers; and 3 runs per week for NextSeq[™] sequencer

3. Ion AmpliSeq[™] library chemistry is automated on Ion Chef[™] instrument

4. Field support provided to portfolio products

16 For Research Use Only. Not for use in diagnostic procedures.

ThermoFisher SCIENTIFIC

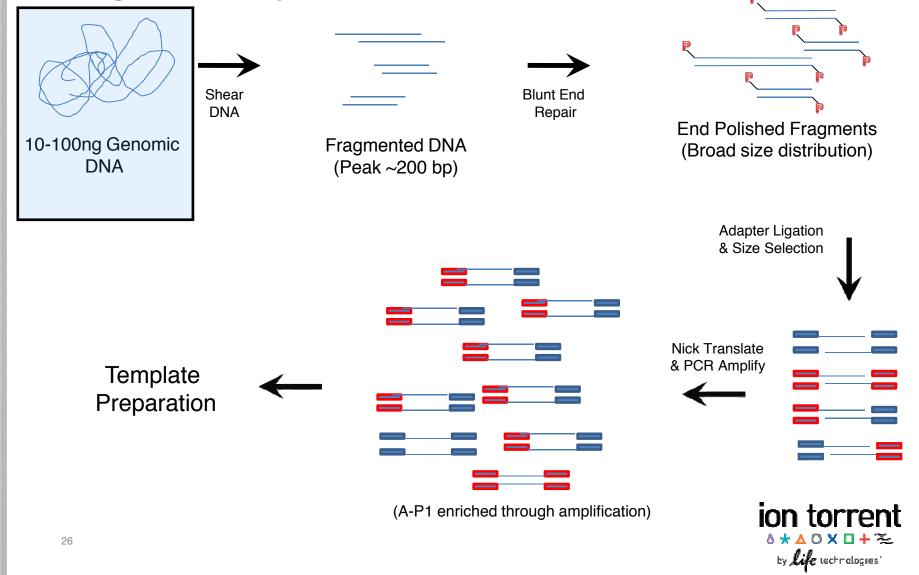
Bridge PCR amplification

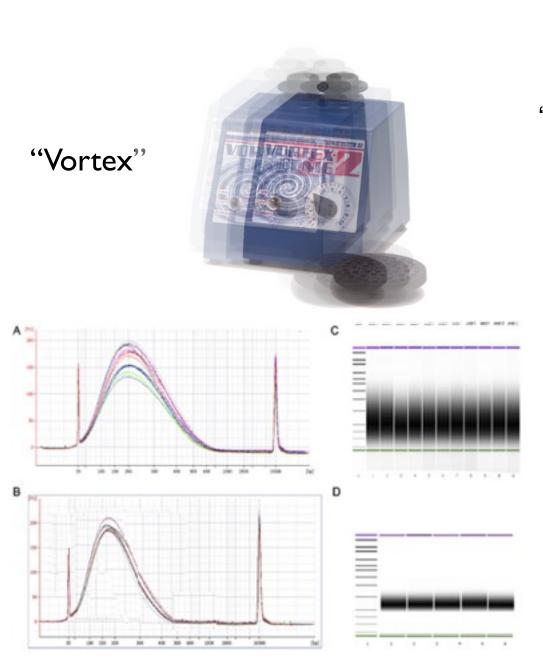
Illumina Sequence Analysis



Ion Proton Next-Gen, Deep Sequencing (ABI/Life technologies)

Ion Fragment Library Kit Fragment library Workflow

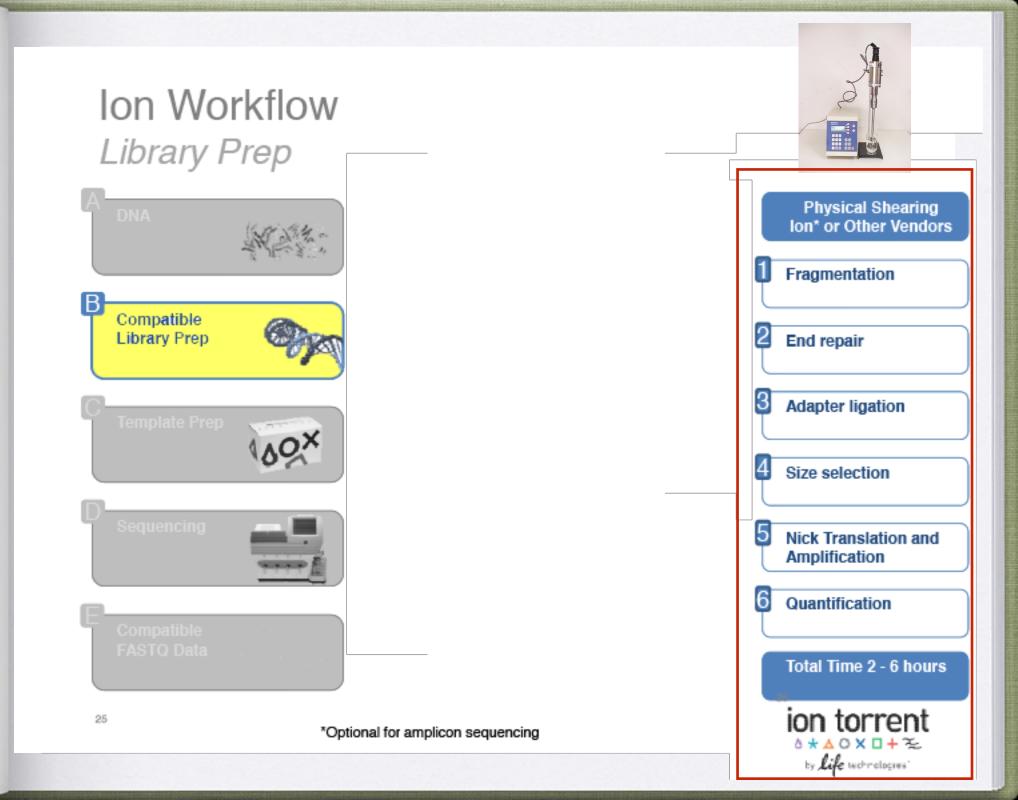


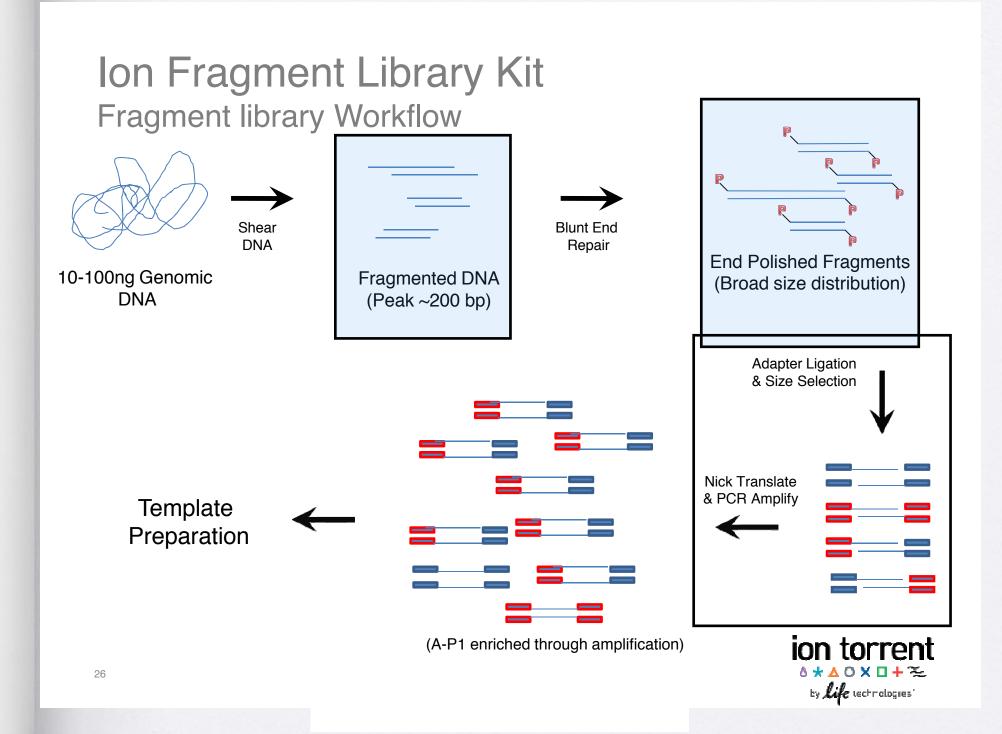


"Sonicator"

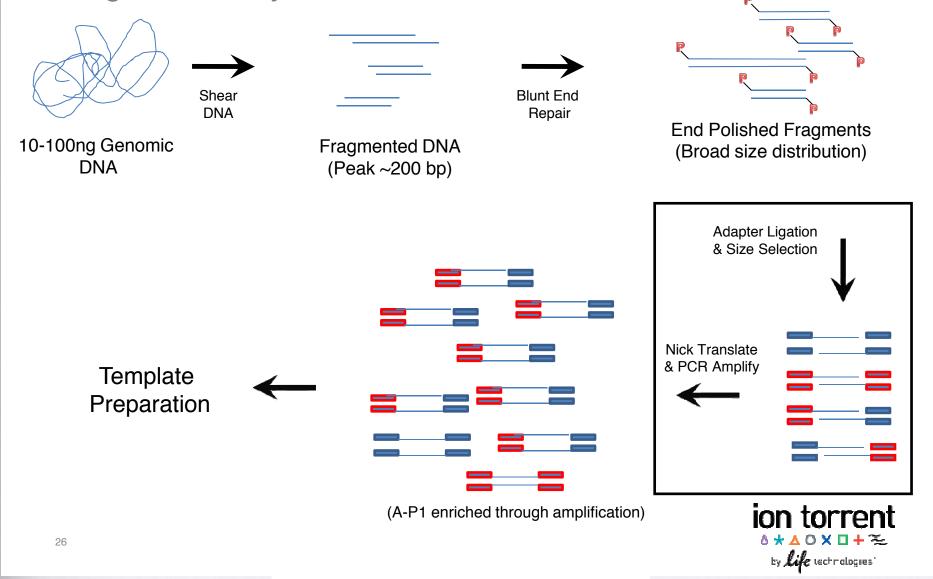
"Shear"

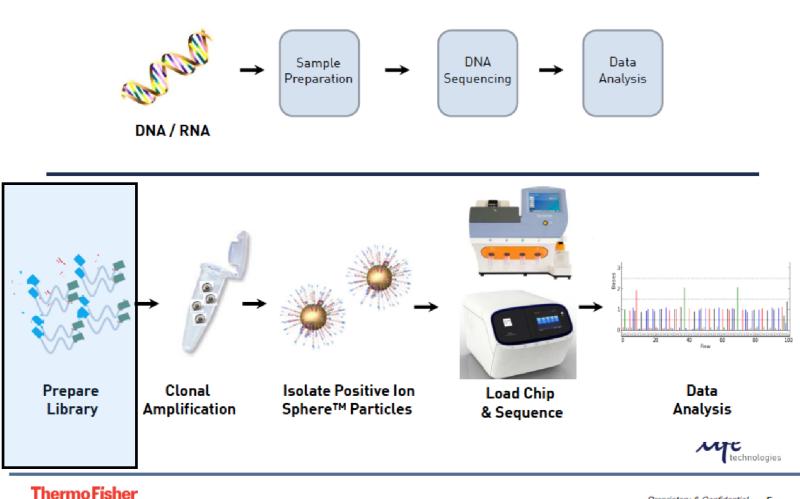




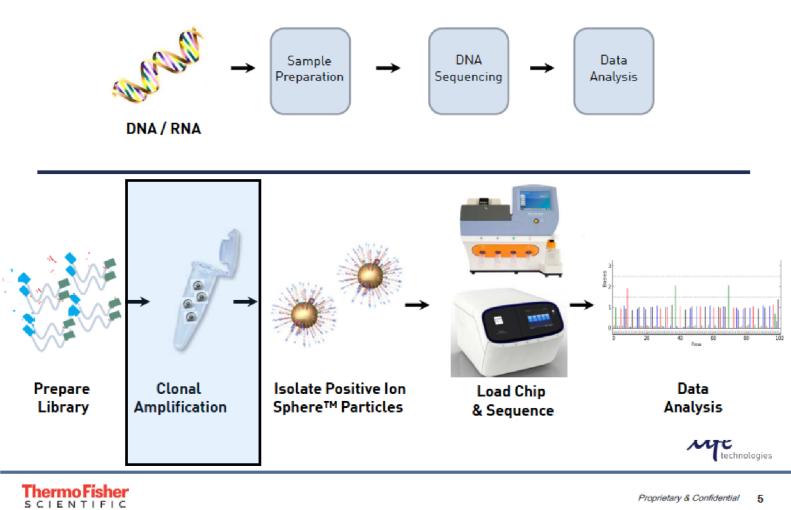


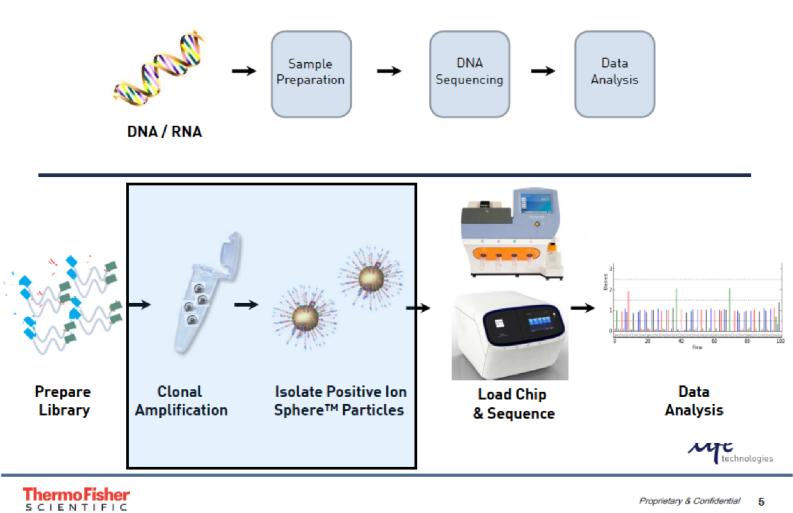
Ion Fragment Library Kit Fragment library Workflow

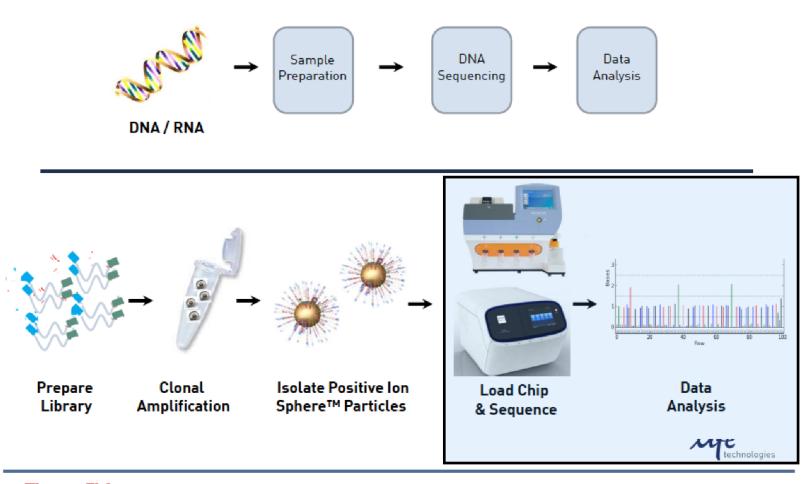




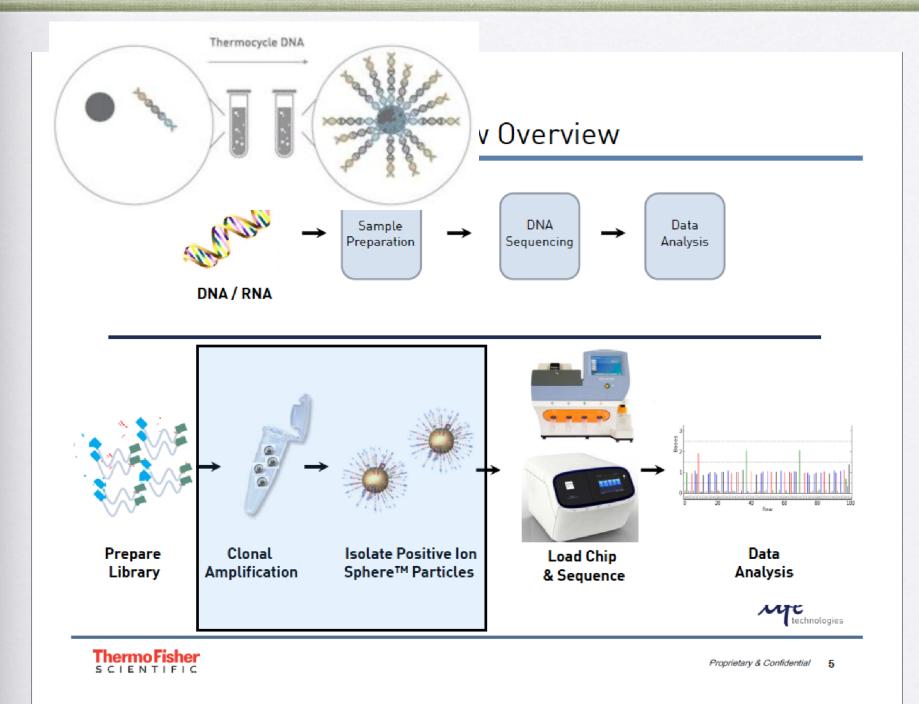
Thermo Fisher

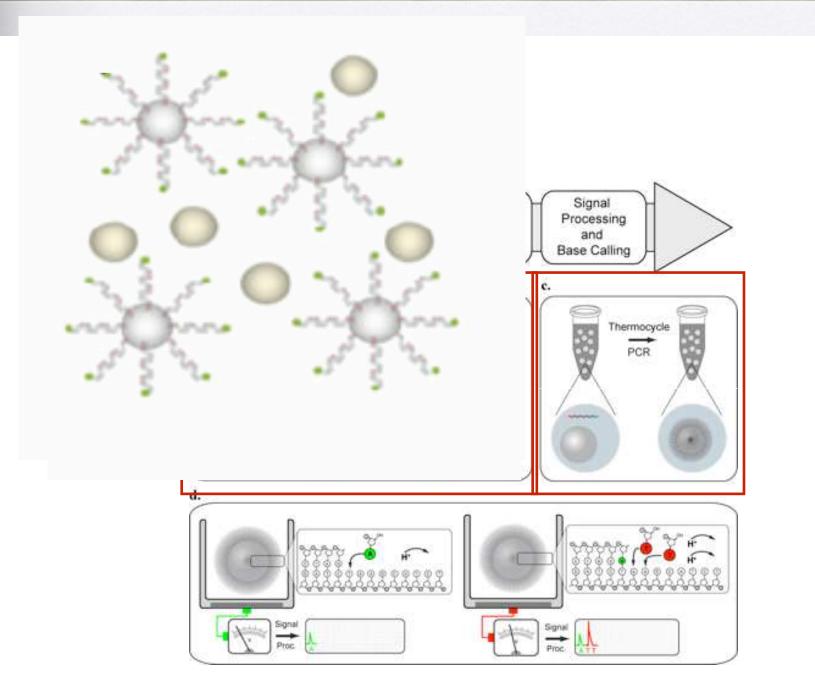




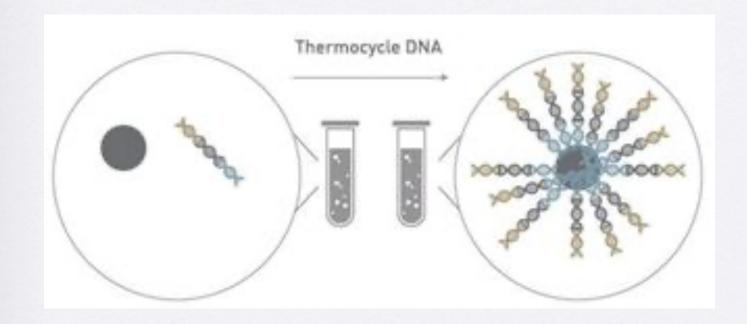


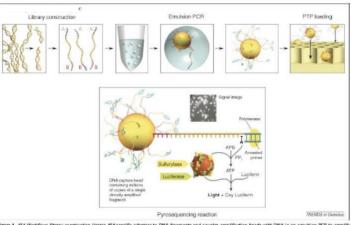
Thermo Fisher



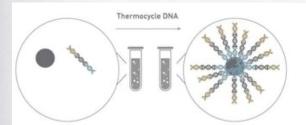


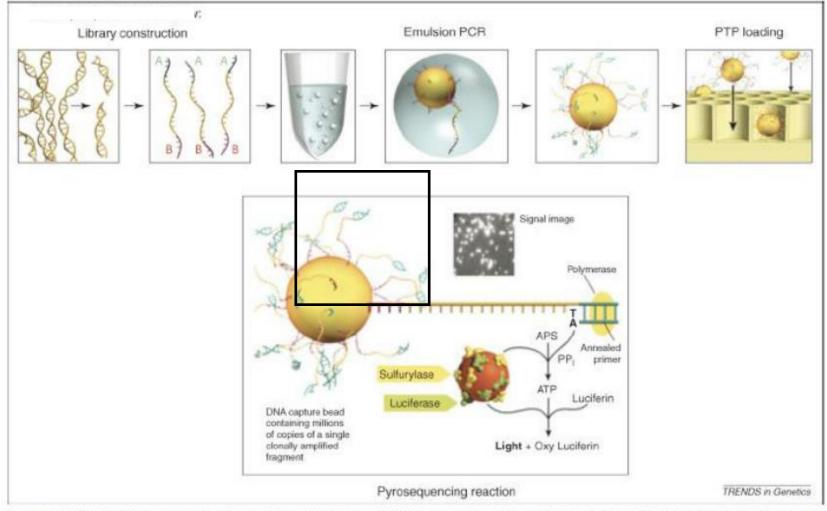
Confidential and Proprietary-DO NOT DUPLICATE





Hgure 1. 454 Workflow: liberry construction lightes 454-specific adapters to ONA fragments and couples amplification beeds with DNA in an emulsion PCR to amplify tragments before sequencing. The basids are leaded into the picotter plate (PTP). The bottom panel illustrates the prosequencing meetors that occurs on nucleotide incorporation to emport sequencing by synthesis.



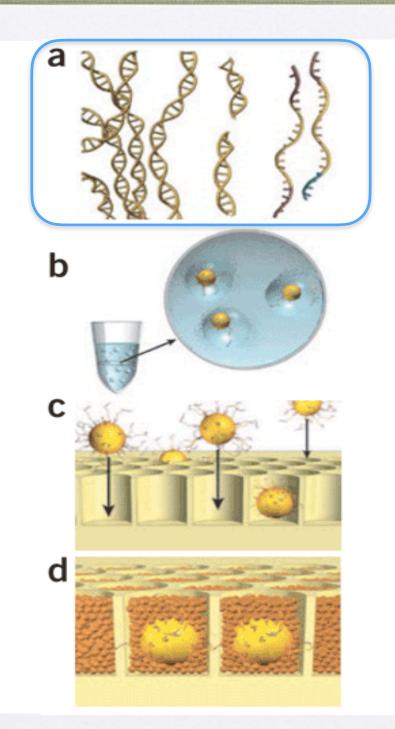


Rgure 1. 454 Workflow: library construction ligates 454-specific adapters to DNA fragments and couples amplification beads with DNA in an emulsion PCR to amplify fragments before sequencing. The beads are loaded into the picotiter plate (PTP). The bottom panel illustrates the pyrosequencing reaction that occurs on nucleotide incorporation to report sequencing by synthesis.

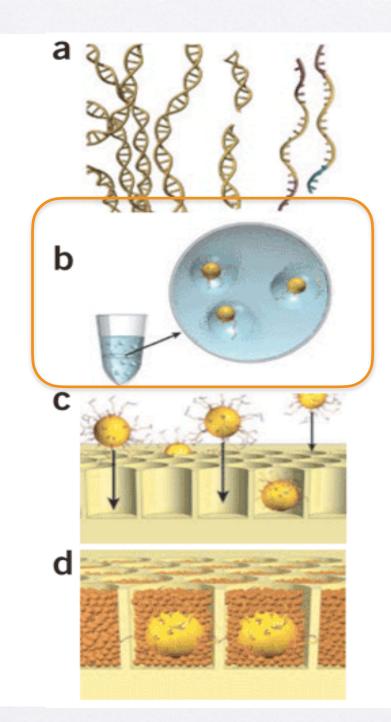


Automation, Ion Chef (ABI/Life technologies)

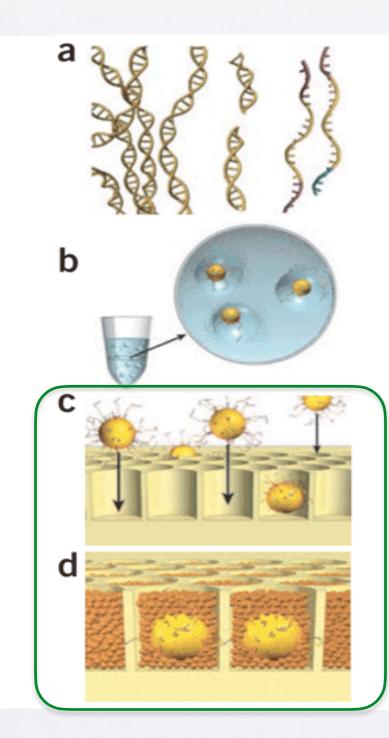








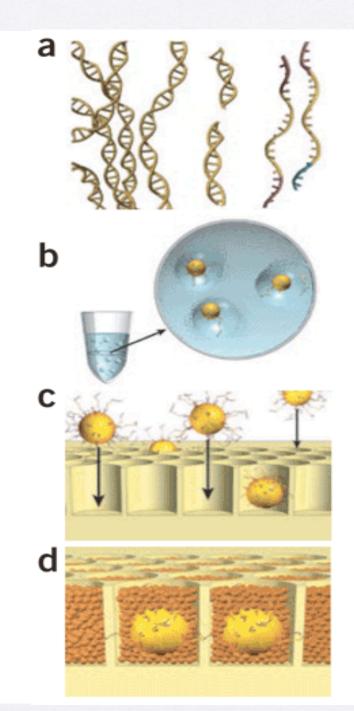


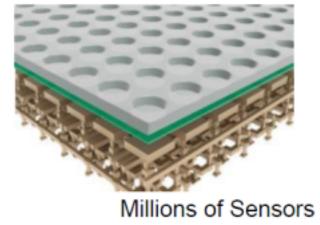




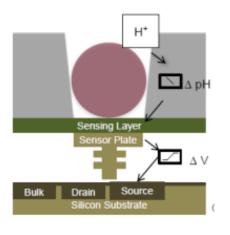




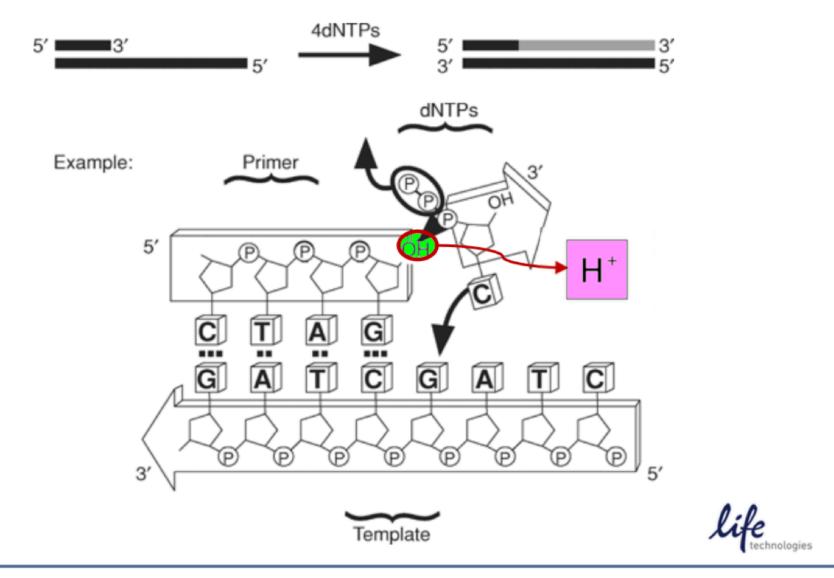




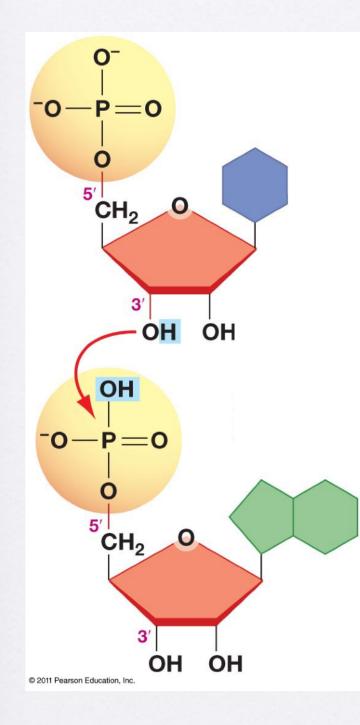
Semiconductor Design



Simple, Natural Chemistry





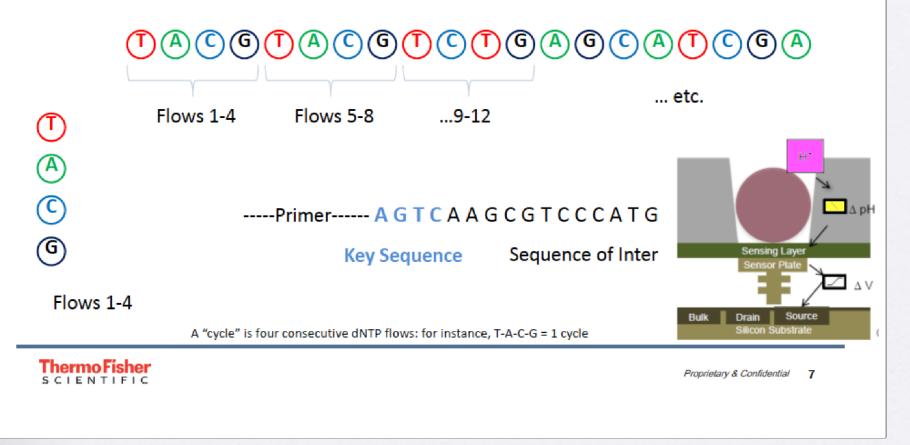


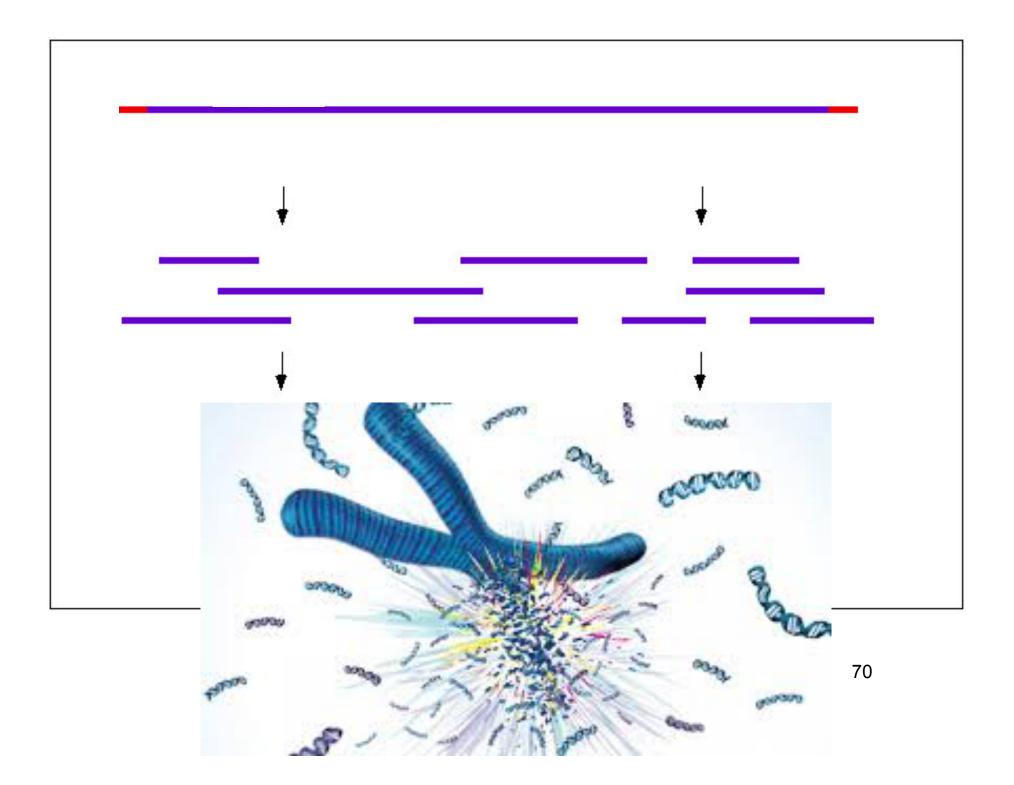
Rapid Direct Signal Detection

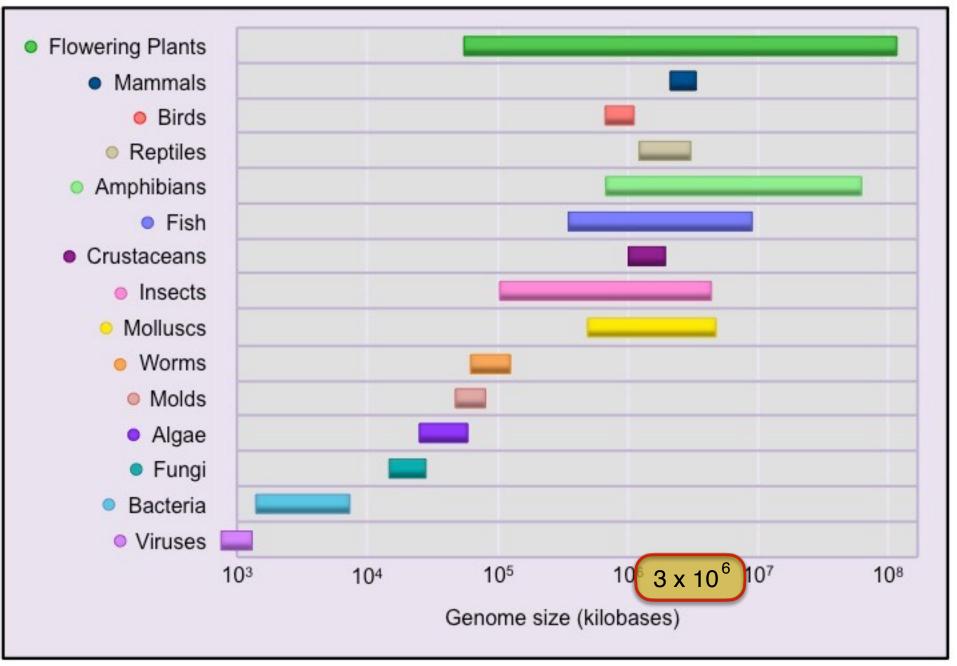


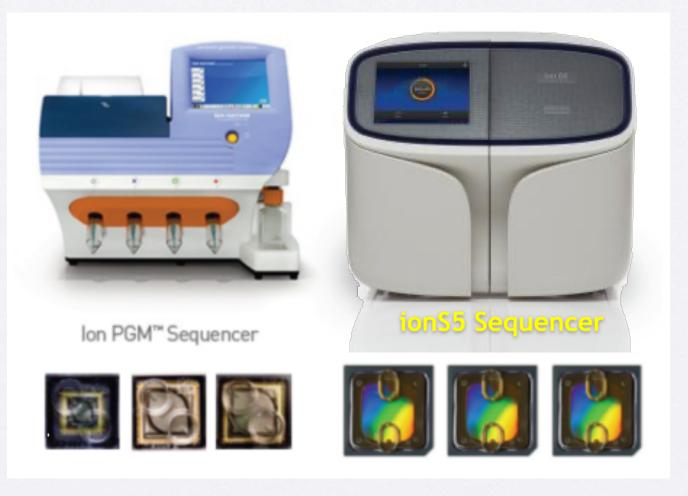
Sequencing: Flows

- A "flow" is the event of exposing the chip to one particular dNTP (T, A, C, or G), followed by a washing step
- The flow order repeats with pattern:
 - 'TACGTACGTCTGAGCATCGATCGATGTACAGC'









	Ion Torrent PGM			Ion Torrent Proton	
Chip Type	PGM 314	PGM 316	PGM 318	Proton I	ionS5
# of sensors	1.3M	6.3M	11M	165M	660M
Total output	10-40Mb	100-400Mb	~1Gb	~10Gb	~100Gb
Run time	1-2 hrs	1-2 hrs	1-2 hrs	2.5 hrs	2.5 hrs
Read length	up to 400bp	~200bp	up to 400bp	~200bp	~200bp
Total reads	up to 0.6M	up to 3M	up to 6M	60-80M	240-330M

Covid-19 Genomic Sequence Analysis @GSU

Equipment/Reagents

- ABS IonTorrent S5 with chef for ISP
- 540 chips offer 80 million reads allowing for- 80 samples per chip
- Ion ampliseq technology for building library
 - 2 pools of ~1,200 primers each
 - Fragment the sequence into roughly 200 bp segments
- Ioncode Barcode adaptors
 - Unique beginning / ending sequences allow for multiple sequences on the same chip



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Pango Lineage Reports

 Sequences that were >98% complete were run through the Pangolin COVID-19 Lineage Assigner (~200 sequences)

https://pangolin.cog-uk.io/

