SUN	MON	TUE	WED	THU	FRI	
June 26	27	28			31	Jı
	9:00-10:00am Virtual Program Orientation for Summer Institute Online Modality					
July 03	04	05	06	07	08	
	Holiday (Independence Day)	8:30-10:00am -Welcome Reception and Buddy Meet & Greet Event	Free Day	Classes begin! 8:30-11am: BIOL4905 INTRODUCTION 8-10:20pm: Afternoon course	8:30-11am: BIOL4905 DNA PREPARATION 8-10:20pm: Afternoon course	
10	11	12	13	14	15	
	8:30-11am:BIOL4905 PROTEOMICS I 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 PROTEOMICS II 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 PROTEOMICS III 8-10:20pm: Afternoon course	8:30-11am: BIOL4905 RNA PREPARATION 8-10:20pm: Afternoon course	Virtual Independence Day Activity	
17	18	19	20	21	22	
	8:30-11am:BIOL4905 qPCR / ROBOTS 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 DNA Sequence Analysis 8-10:20pm: Afternoon course	Midterm Break	8:30-11am:BIOL4905 Next Gen. Sequencing 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 Automated Microscopy /AFM	
24	25	26	27	28	29	
	8:30-11am:BIOL4905 Microarray I 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 Microarray II 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 Nanostring 8-10:20pm: Afternoon course	8:30-11am:BIOL4905 Flow Cytometry 8-10:20pm: Afternoon course	FINALS	
31	August 01	02	03			
	9:00-10:00am: Closing Reception		Grades available in PAWS			



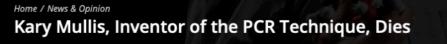


### Fundamentals of Real-Time RT-PCR

adapted from a PPT presentation by David Chappell, PhD ABI Field applications Specialist



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NEWS & OPINION	MAGAZINE	SUBJECTS	MULTIMEDIA	CAREERS		



The Nobel laureate was a proponent of LSD, a consultant for O.J. Simpson's legal defense, and the creator of a company that infused jewelry with celebrities' DNA.



🗟 🛃 🕇 🏹 🕂 47K

**K** ary Mullis, whose invention of the polymerase chain reaction technique earned him the Nobel Prize in Chemistry in 1993, died of pneumonia on August 7, according to <u>MyNewsLA.com</u>. He was 74 years old.

ABOVE: FLICKR, ERIK CHARLTON

According to a 1998 profile in *The Washington Post*, Mullis was known as a "weird" figure in science and "flamboyant" philanderer who evangelized the use of LSD, denied the evidence for both global warming and HIV as a cause of AIDS, consulted for O.J. Simpson's legal defense, and formed a company that sold jewelry embedded with celebrities' DNA. The opening paragraph of his Nobel autobiography includes a scene depicting a visit from Mullis's dying grandfather in "non-substantial form."

"He was personally and professionally one of the more iconic personalities science has ever witnessed," Rich Robbins, the founder and CEO of Wareham Development, a real estate developer for a number of biotech companies, tells the Emeryville, California-based paper, the *E'ville Eye*.

#### See "PCR: Past, Present, & Future"

Mullis was born in North Carolina in 1944 and earned a chemistry degree from Georgia Tech and a PhD in biochemistry from the University of California, Berkeley. In the early 1980s, when Mullis



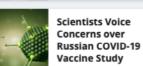
x Q & Login

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#### Trending







(1) Denaturation 5' Annealing 3) Elongatior + + 2&3 ,2&3 1,2&3

5

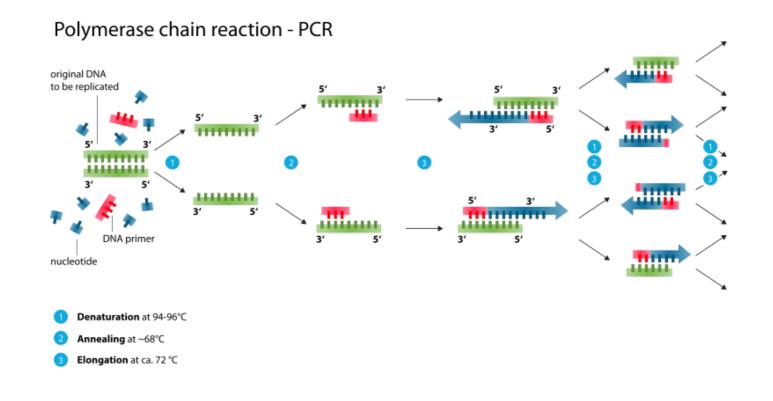
Exponential growth of short product

#### PCR and other inventions

Main articles:

#### and

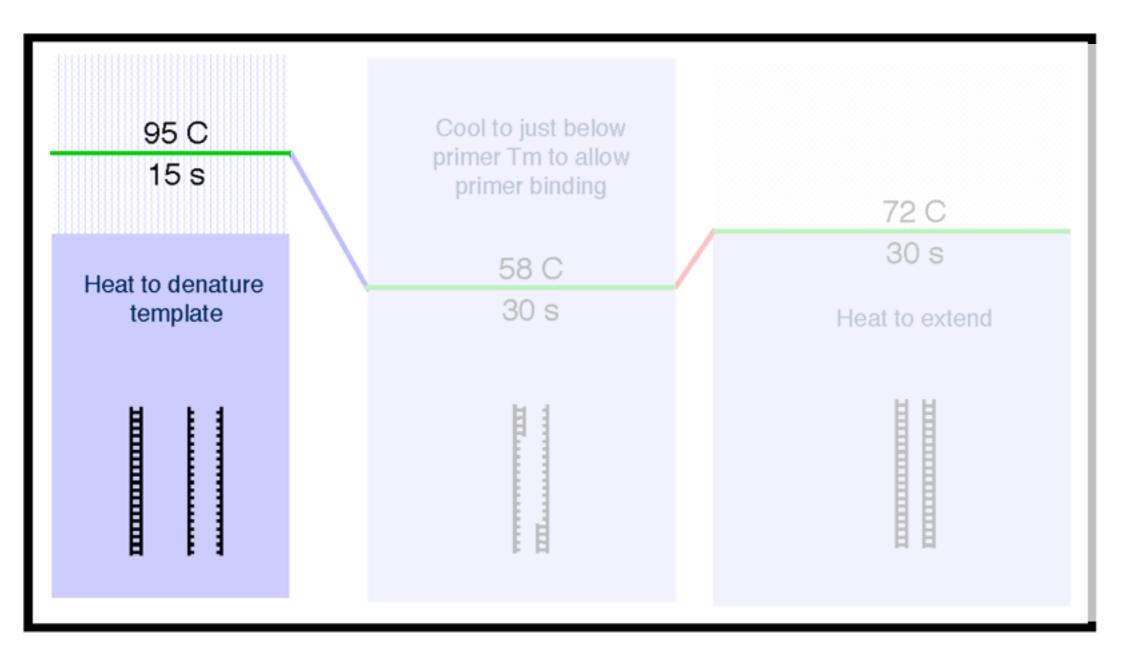
In 1983, Mullis was working for Cetus Corporation as a chemist. Late one night while driving with his girlfriend, who was also a chemist at Cetus, he had the idea to use a pair of primers to bracket the desired DNA sequence and to copy it using DNA polymerase; a technique that would allow rapid amplification of a small stretch of DNA and become a standard procedure in molecular biology laboratories.<sup>[10]</sup> Cetus took Mullis off his usual projects to concentrate on PCR full-time. Mullis succeeded in demonstrating PCR December 16, 1983.<sup>[10]</sup> He received a \$10,000 bonus from Cetus for the invention.



A drawback of the technique was that the DNA polymerase in the reaction was destroyed by the high heat used at the start of each replication cycle and had to be replaced. In 1986, Saiki started to use *Thermophilus aquaticus* (Taq) DNA polymerase to amplify segments of DNA. The Taq polymerase was heat resistant and only need to be added to the reaction once, making the technique dramatically more affordable and subject to automation. This modification of Mullis' invention revolutionized biochemistry, molecular biology, genetics, medicine, and forensics.



ycles



#### Traditional PCR





1990 Microcycler: Eppendorf introduces its first thermal cycler using water to heat and cool.

1993 Mastercycler 5330: Eppendorf introduces the first Mastercycler based on peltier technology.



2005

Mastercycler ep regipiex: Extremely fast optics for rapid data acquisition.



### 2008

Mastercycler pro: New vapa.protect\*\* technology reduces evaporation.

Temperature control range of the block	4–99 °C					
Temperature control mode	Fast, Standard, Safe					
Heating technology of the block	Peltier elements, Triple Circuit Technology					
Gradient block	over 12	over 24 columns				
Gradient range	1-20 °C	1-24 °C	1-20 °C			
Gradient temperature range		30-99 °C				
Lid temperature range		37–110 °C				
Lid descent and clos- ing pressure	vapo.protect <sup>™</sup> technology with Thermal Sample Protection					
Block homogeneity: 20 °C–72 °C 95 °C	≤ ±0.3 °C ≤ ±0.4 °C					
Block temperature accuracy		± 0.2 °C				
Heating rate*	ca. 4 °C/s	ca. 6 °C/s	ca. 4 °C/s			
Cooling rate*	ca. 3 °C/s	ca. 4,5 °C/s	ca. 3 °C/s			
Interfaces	Centronics, USB, CAN in, CAN out					
Dimensions (W × D × H)	26 × 41.5 × 37 cm					
Weight		18.5 kg (40.8 lbs)				
Power supply	230 V, 50-60 Hz					
Max. power consumption	950 W					
Sound power levels	≤ 56 dB(A)					

\*\*Unit can only be operated via a Mastercycler nexus unit (including flat, X1 versions) with control and display panel Product appearance and/or specifications are subject to change without notice.



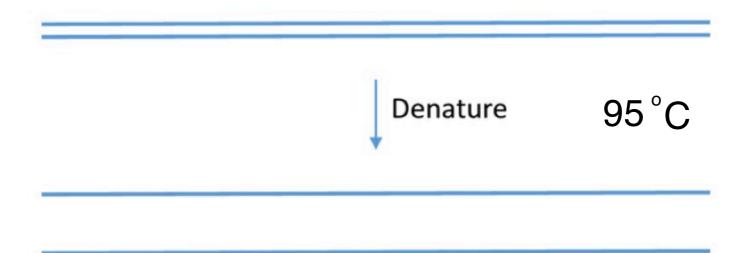
#### Traditional PCR – examine products at the <u>end</u> of the reaction

		Cycle 1	Cycle 2	2	Cycle 3	Cycle 4		
		<b>→</b>	<b>→</b>		<b>→</b>	<b>→</b>	$\rightarrow \rightarrow \rightarrow$	
$\overline{\ }$	_	_						



#### Mastercycler Gradient Pro -Thermal Cycler

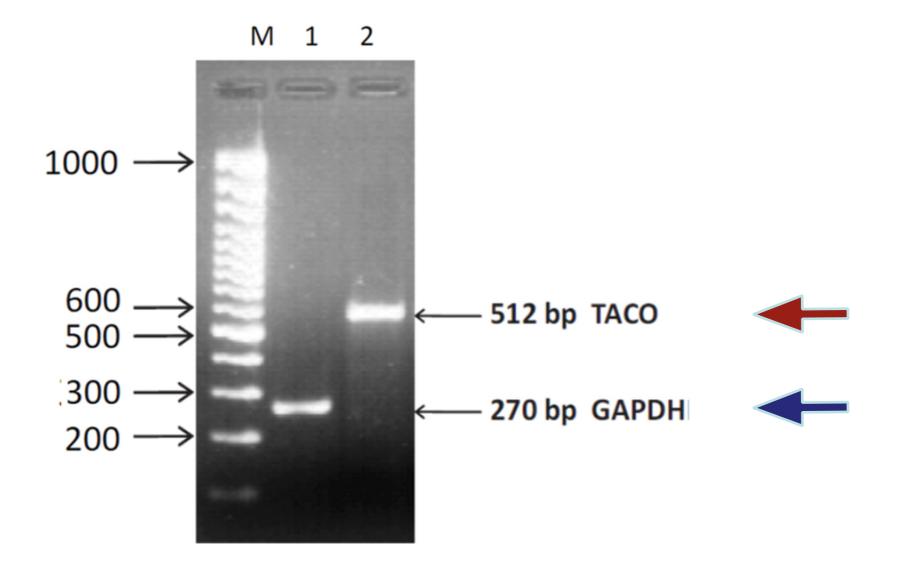
- Major reduction of evaporation in tubes
- Extremely fast heating and cooling rates
- Gradient blocks with SteadySlope technology
- Intuitive graphic programming
- Display to indicate cycler number in a network
- Optional self-test of peltier elements

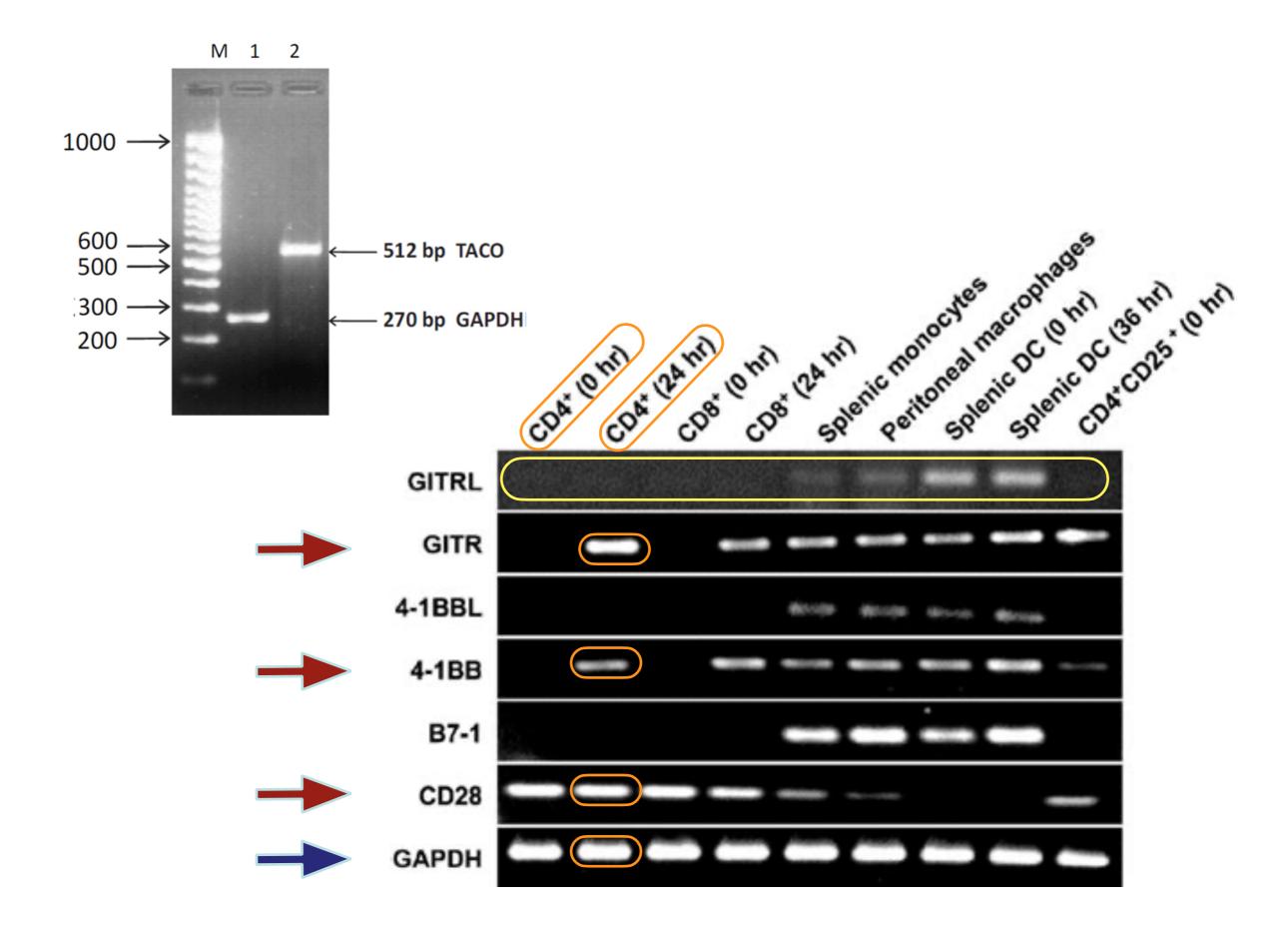




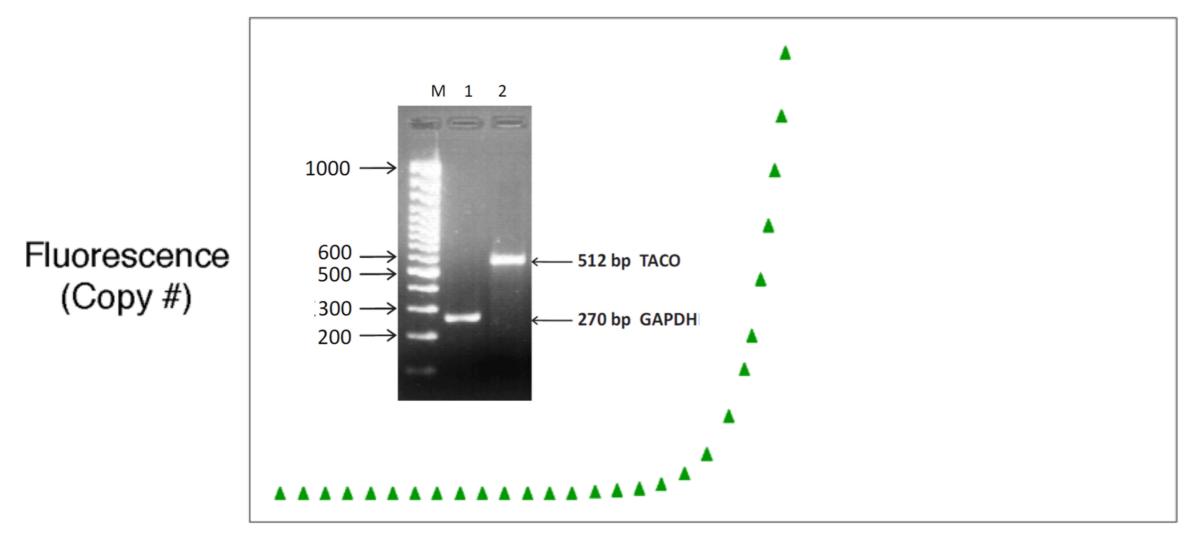
20.





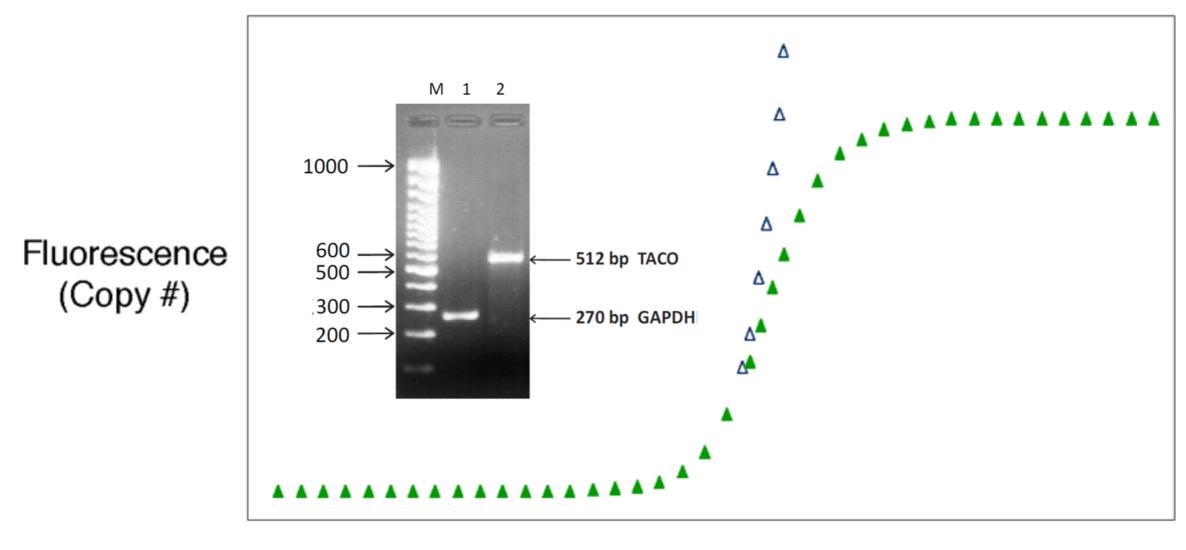






Cycle #





Cycle #



## Real-time PCR or qPCR

# SYBR<sup>®</sup> Green

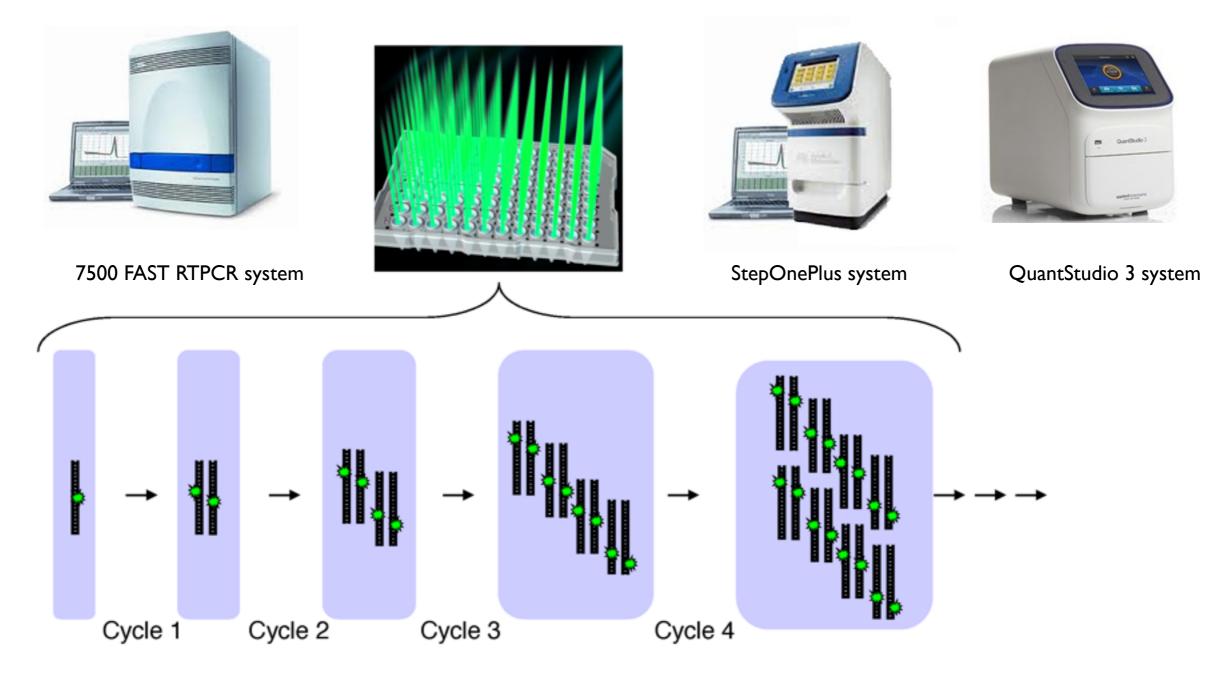
# TaqMan<sup>®</sup>

## MGB

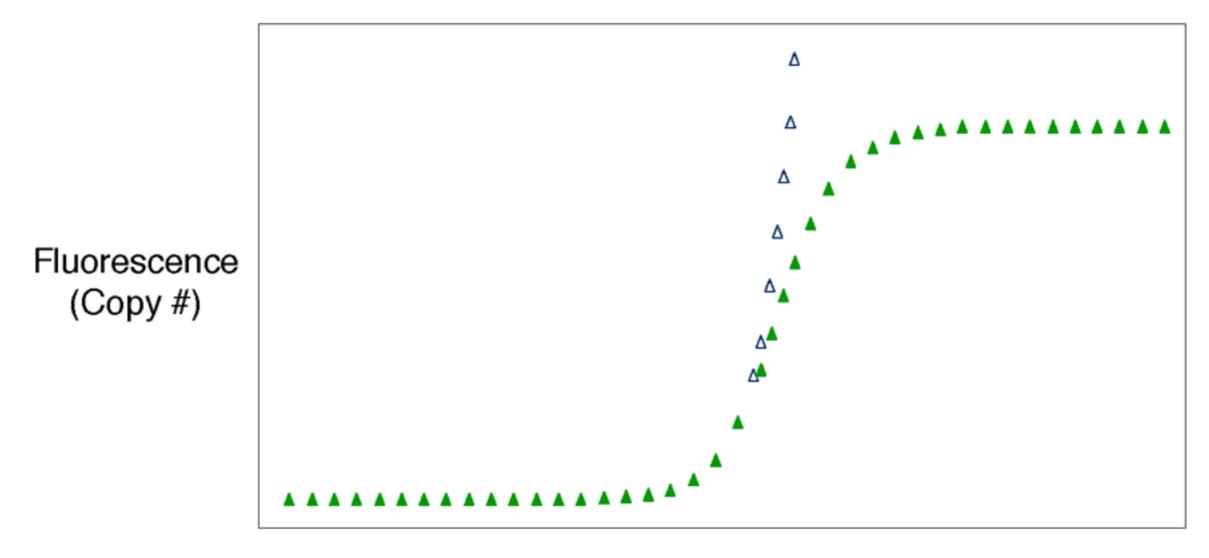
ROX<sup>™</sup>

# Multicomponenting



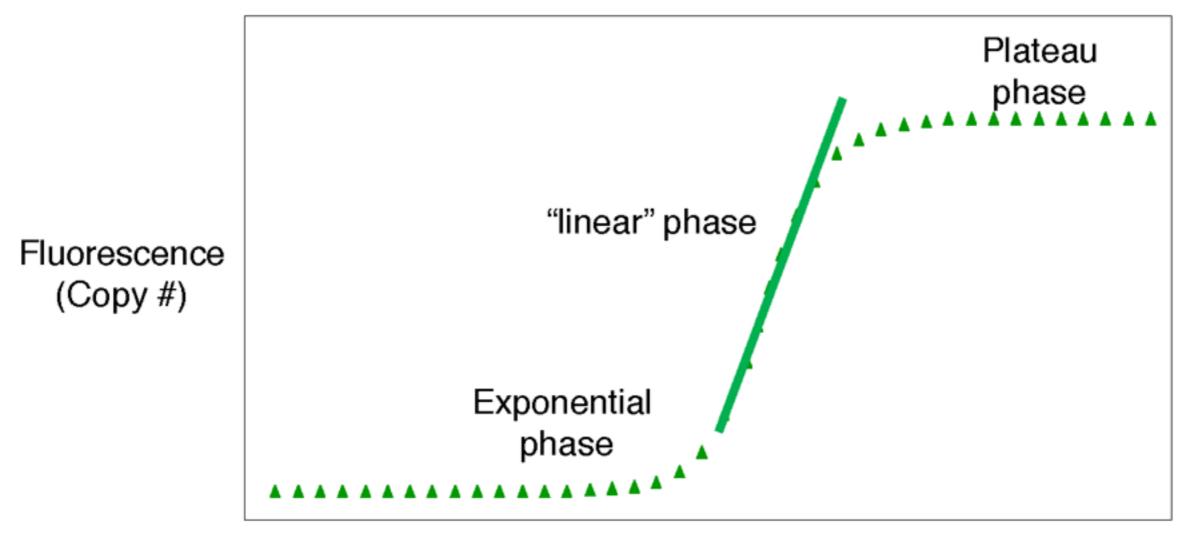






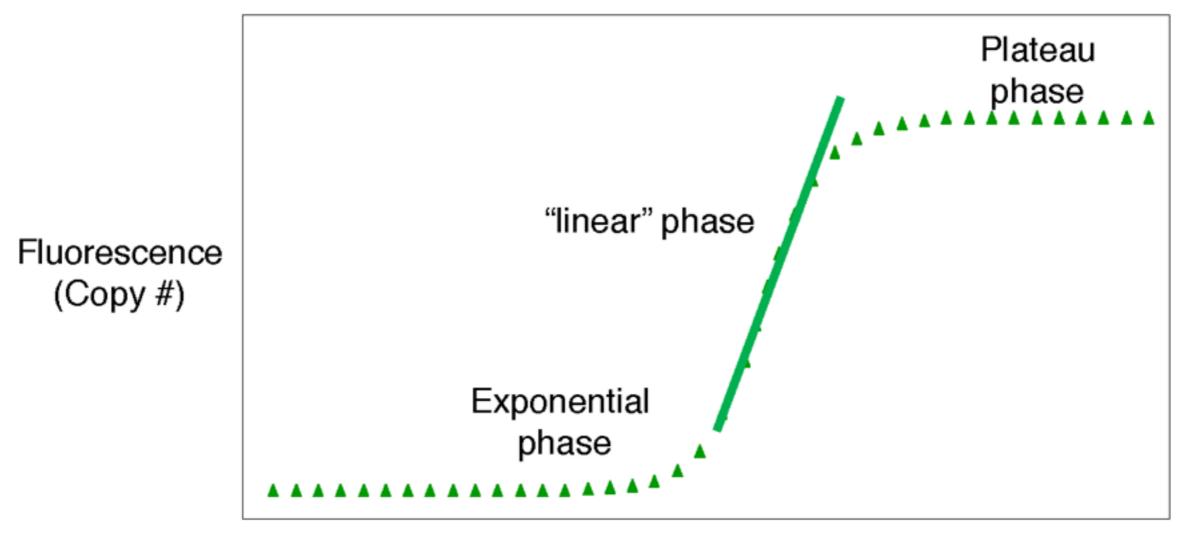
Cycle #





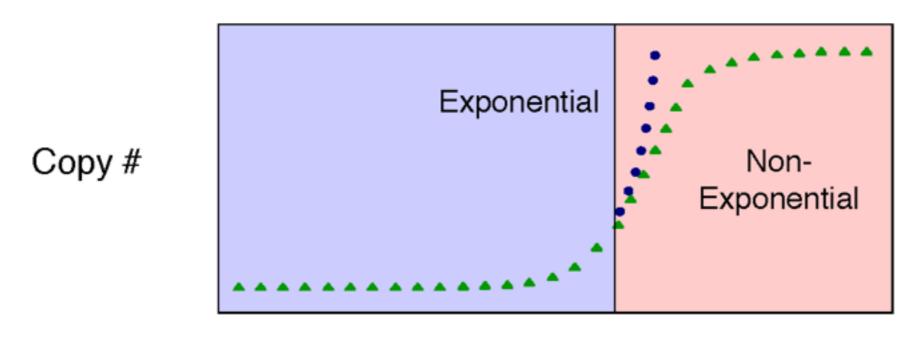
Cycle #





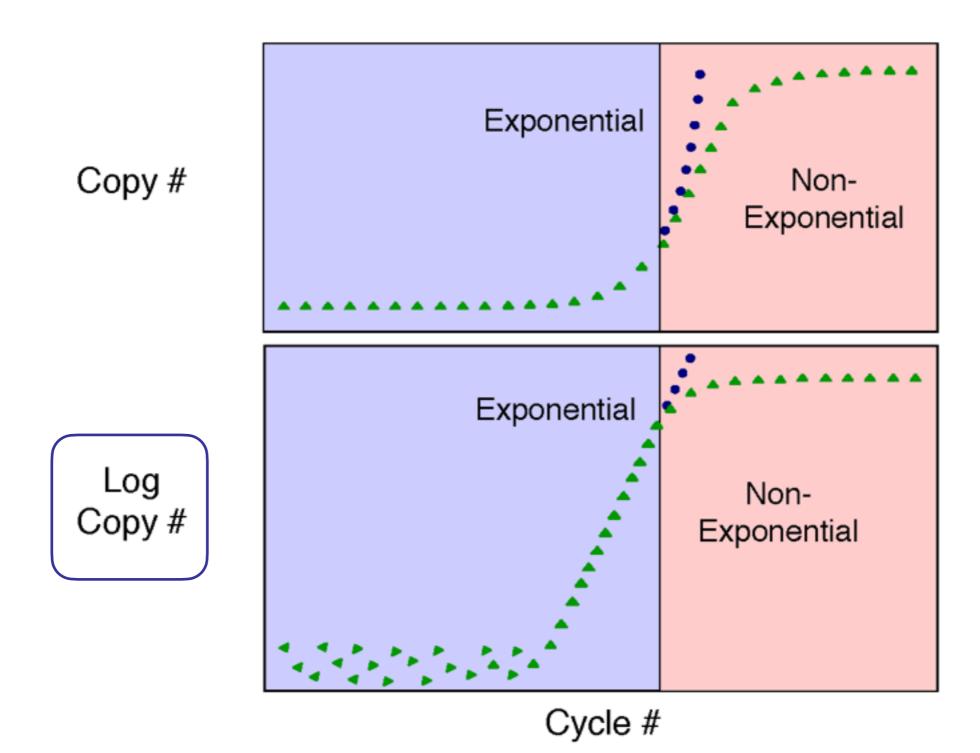
Cycle #





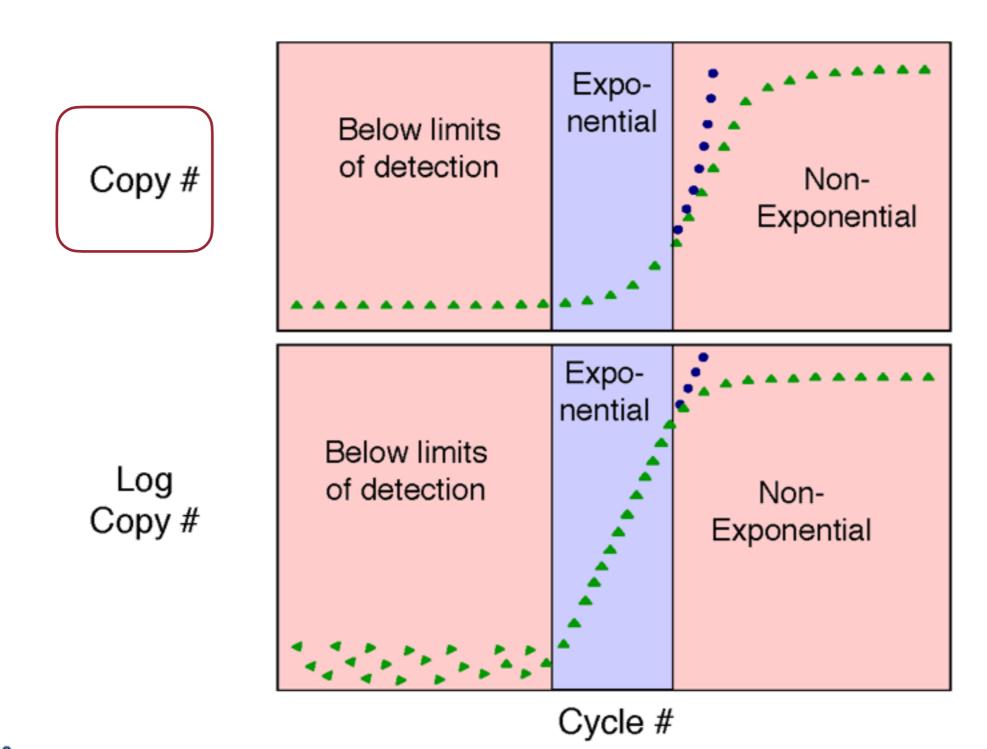
Cycle #





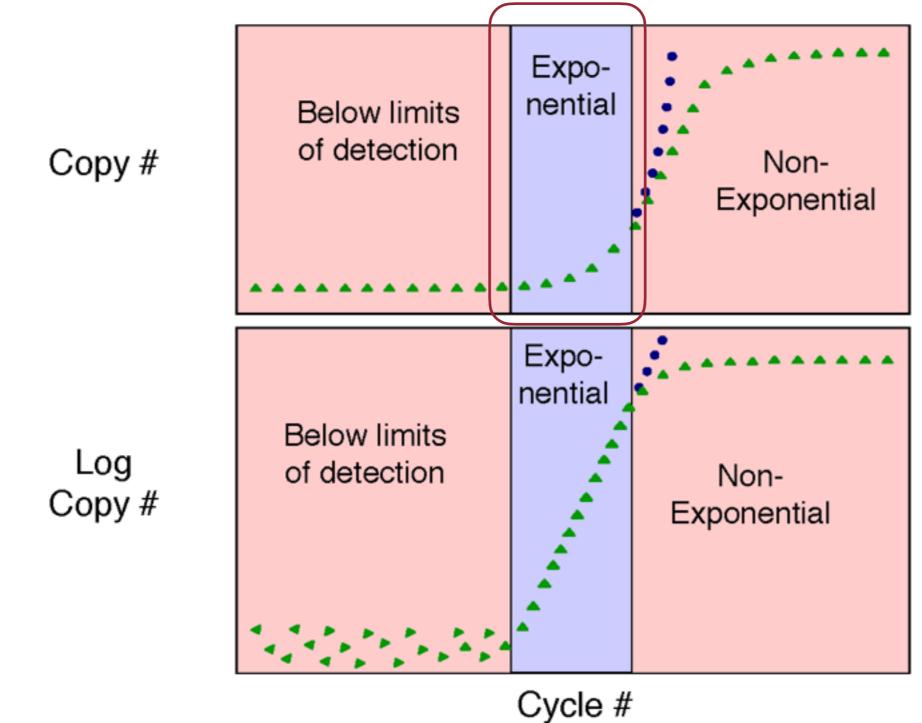


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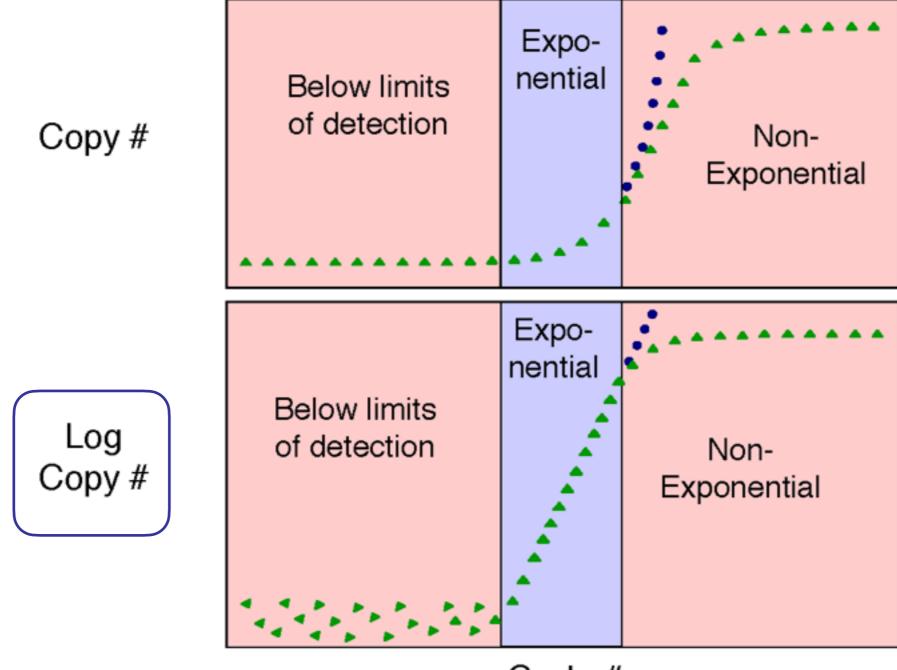


#### The exponential region is easier to define in log phase



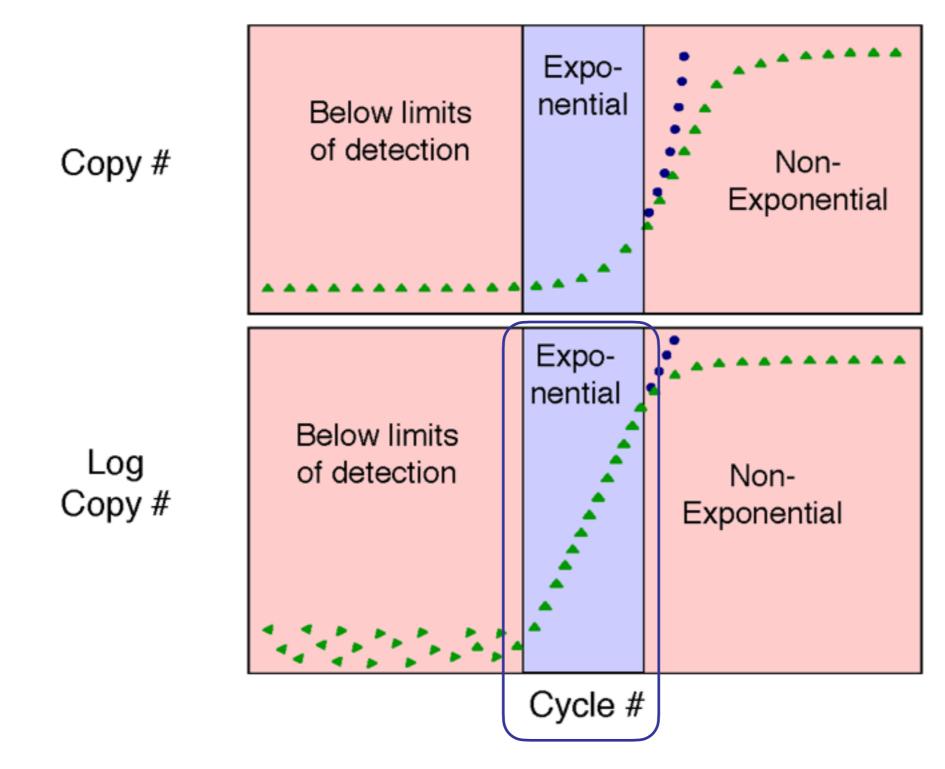


#### The exponential region is easier to define in log phase



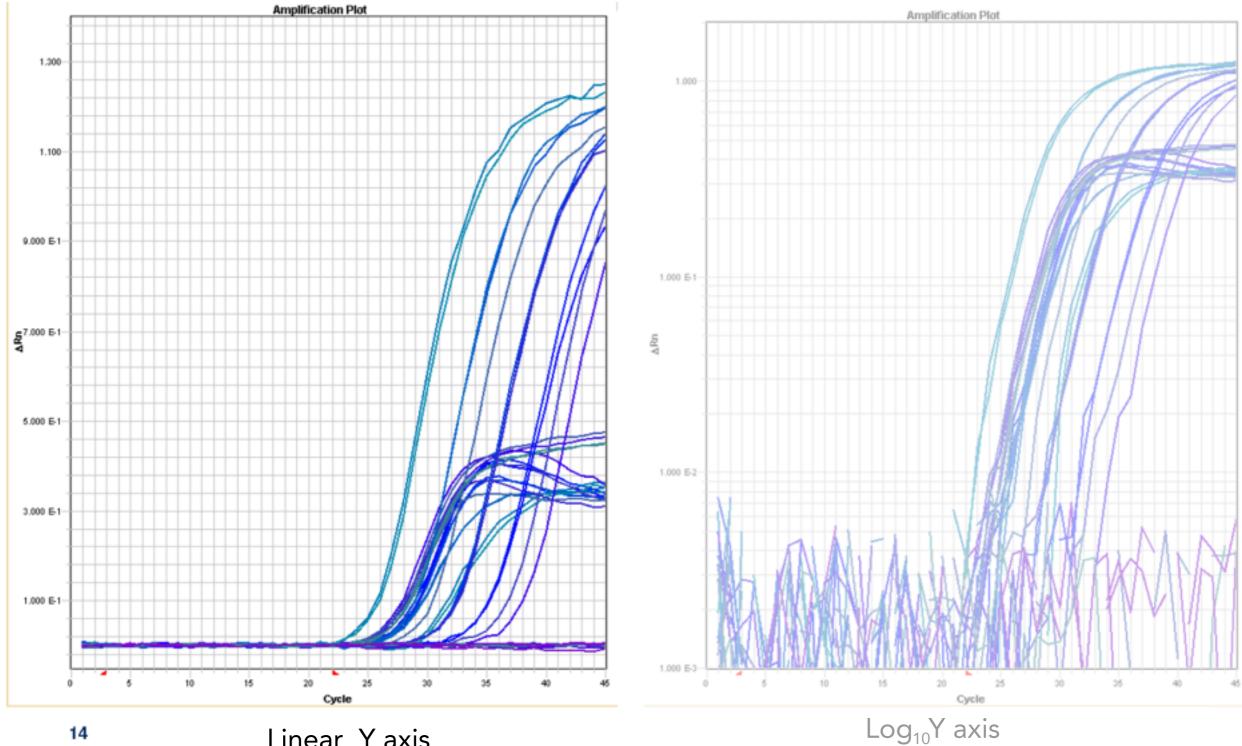


#### The exponential region is easier to define in log phase



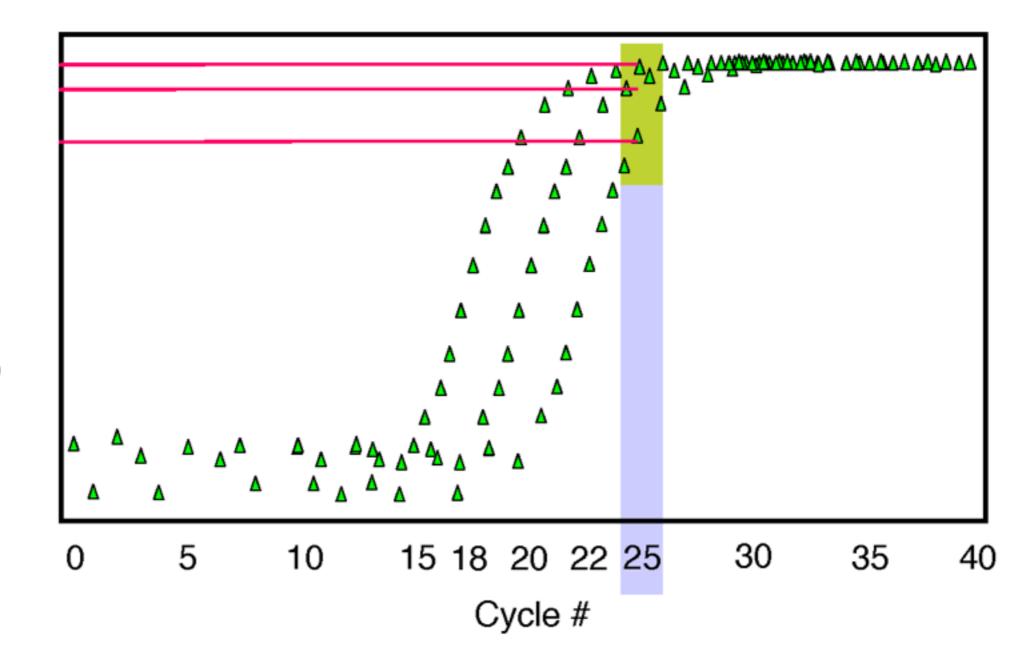


#### Linear and Log view of the same data



Linear Y axis

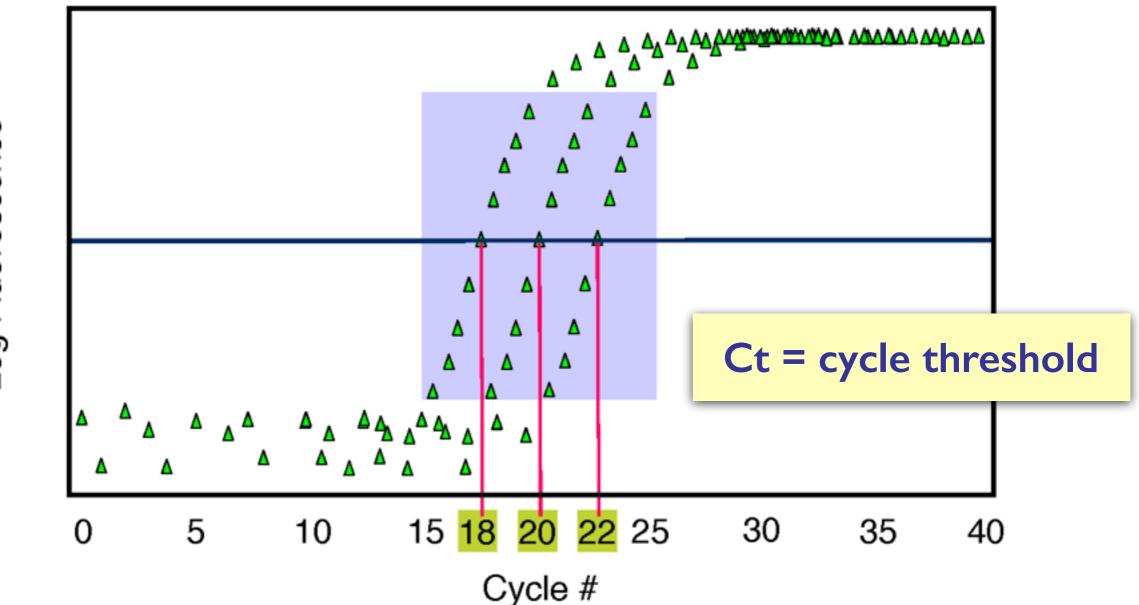




Log Fluorescence

15

#### Real-time PCR - Concept of Ct We measure the number of cycles it takes to reach a set fluorescence threshold (Ct)



Log Fluorescence

16

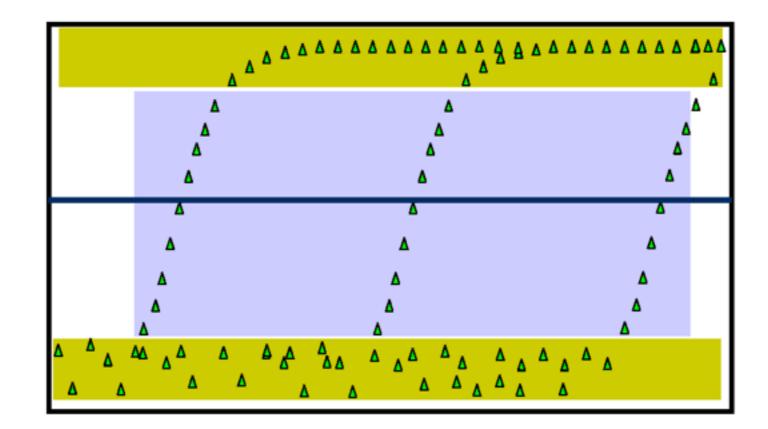


### Thus, real-time PCR is superior to regular PCR because:

	Δ Δ Δ Δ Δ						



#### Thus, real-time PCR is superior to regular PCR because:

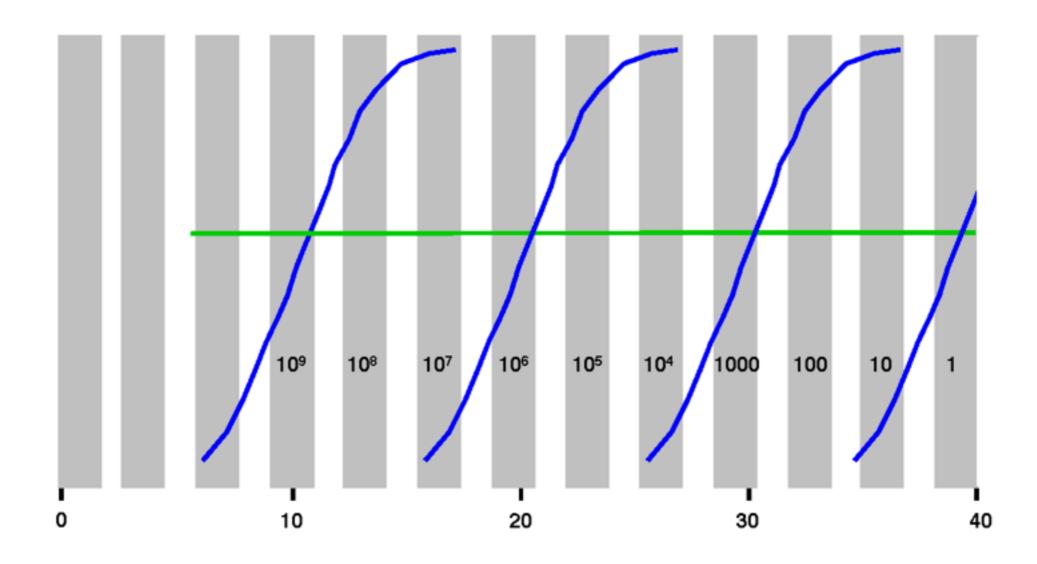


### Results are usually given in table form indicating Ct value

#### the higher the Ct, the lower the target copy number

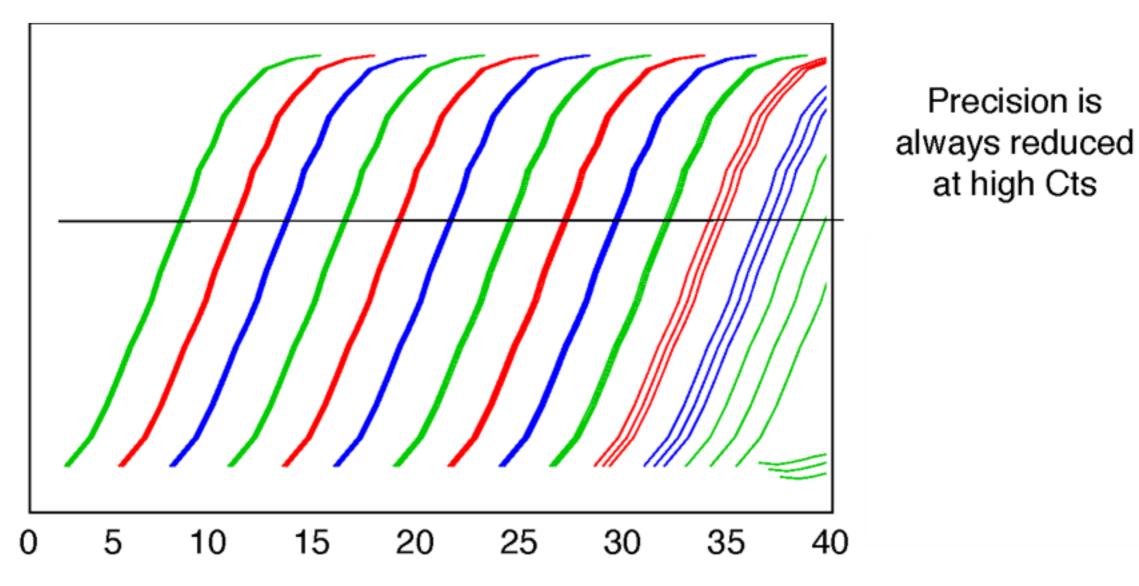
Well	Detector	ng RNA	Ct	Well	Detector	ng RNA	Ct
A4	GAPDH 1	40	16.75	A1	TNF-a 1	40	27.32
A5	GAPDH 1	40	16.89	A2	TNF-a 1	40	27.34
A6	GAPDH 1	40	16.86	A3	TNF-a 1	40	27.28
B4	GAPDH 1	4	20.27	B1	TNF-a 1	4	30.83
B5	GAPDH 1	4	20.24	B2	TNF-a 1	4	30.91
B6	GAPDH 1	4	20.24	B3	TNF-a 1	4	30.87
C4	GAPDH 1	0.4	23.75	C1	TNF-a 1	0.4	34.13
C5	GAPDH 1	0.4	23.71	C2	TNF-a 1	0.4	34.32
C6	GAPDH 1	0.4	23.76	C3	TNF-a 1	0.4	34.25
D4	GAPDH 1	0.04	27.21	D1	TNF-a 1	0.04	38.46
D5	GAPDH 1	0.04	27.18	D2	TNF-a 1	0.04	38.42
D6	GAPDH 1	0.04	27.17	D3	TNF-a 1	0.04	37.18
E4	GAPDH 1	0.004	30.46	E1	TNF-a 1	0.004	Undetermined
E5	GAPDH 1	0.004	29.98	E2	TNF-a 1	0.004	Undetermined
E6	GAPDH 1	0.004	30.6	E3	TNF-a 1	0.004	Undetermined
_							





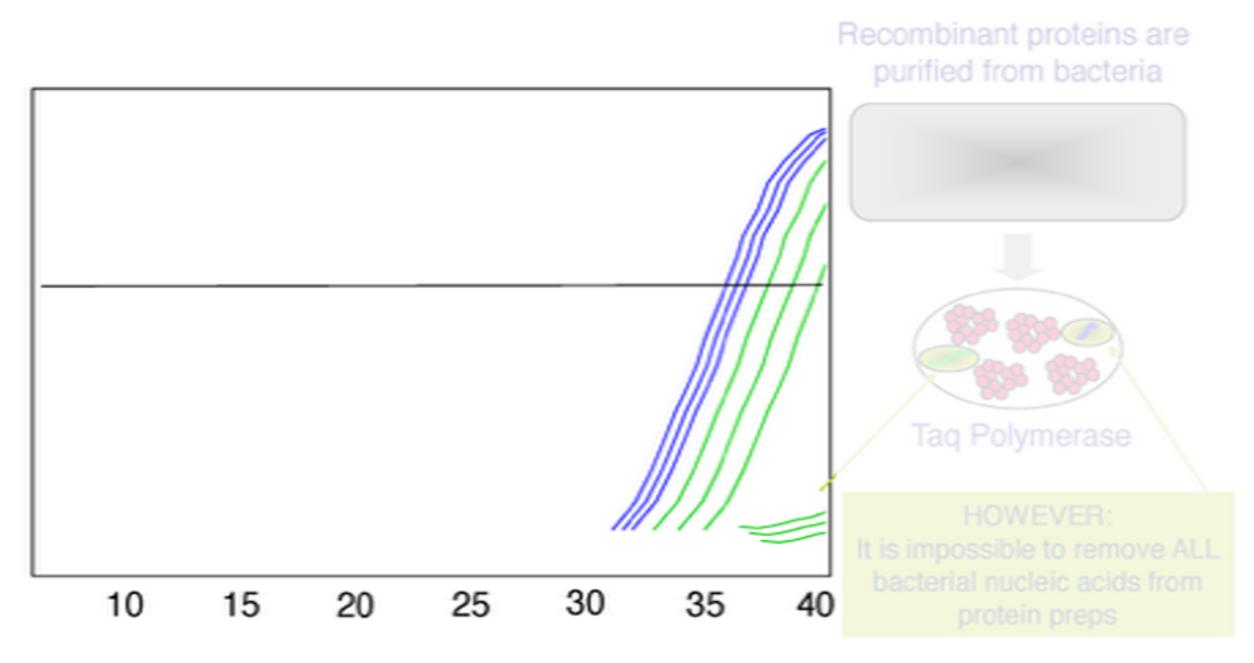


how reproducible is the data?
this is determined by replicates



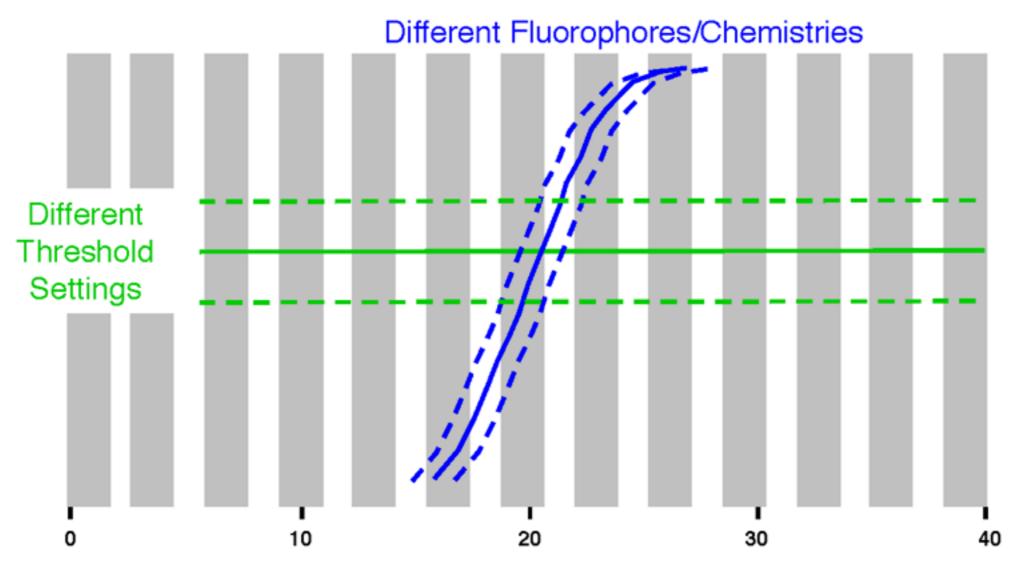


#### NTCs may show some amplification





#### The **Exact** Ct value may vary due to:



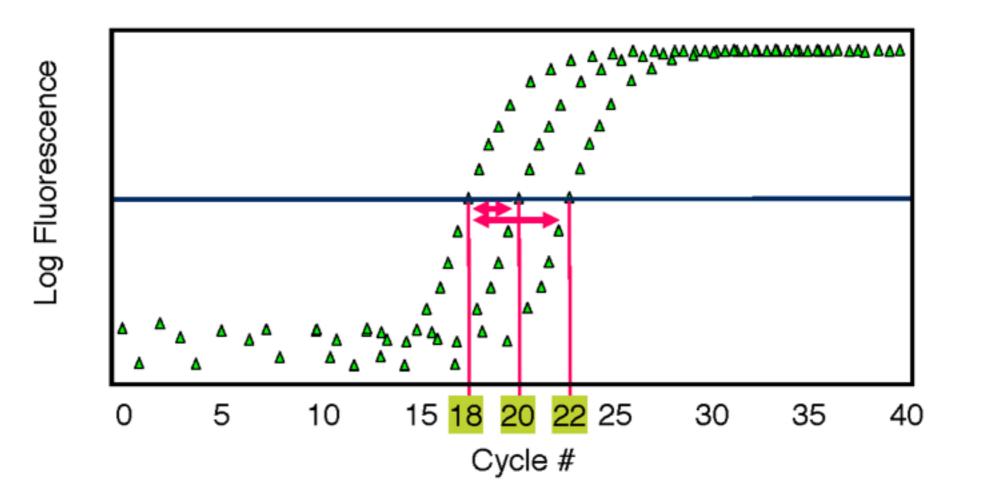
and therefore only indicates an approximate copy number. For this reason, Ct values are not normally published

However, if we compare Cts from the SAME PLATE, then we can be extremely accurate.



### Quantitative real-time PCR analysis measures the DIFFERENCE in the Cts

Either the difference between Sample Cts and Std Cve Cts (Absolute) Or, the difference between sample Cts directly (Relative)





### **Real-time PCR**

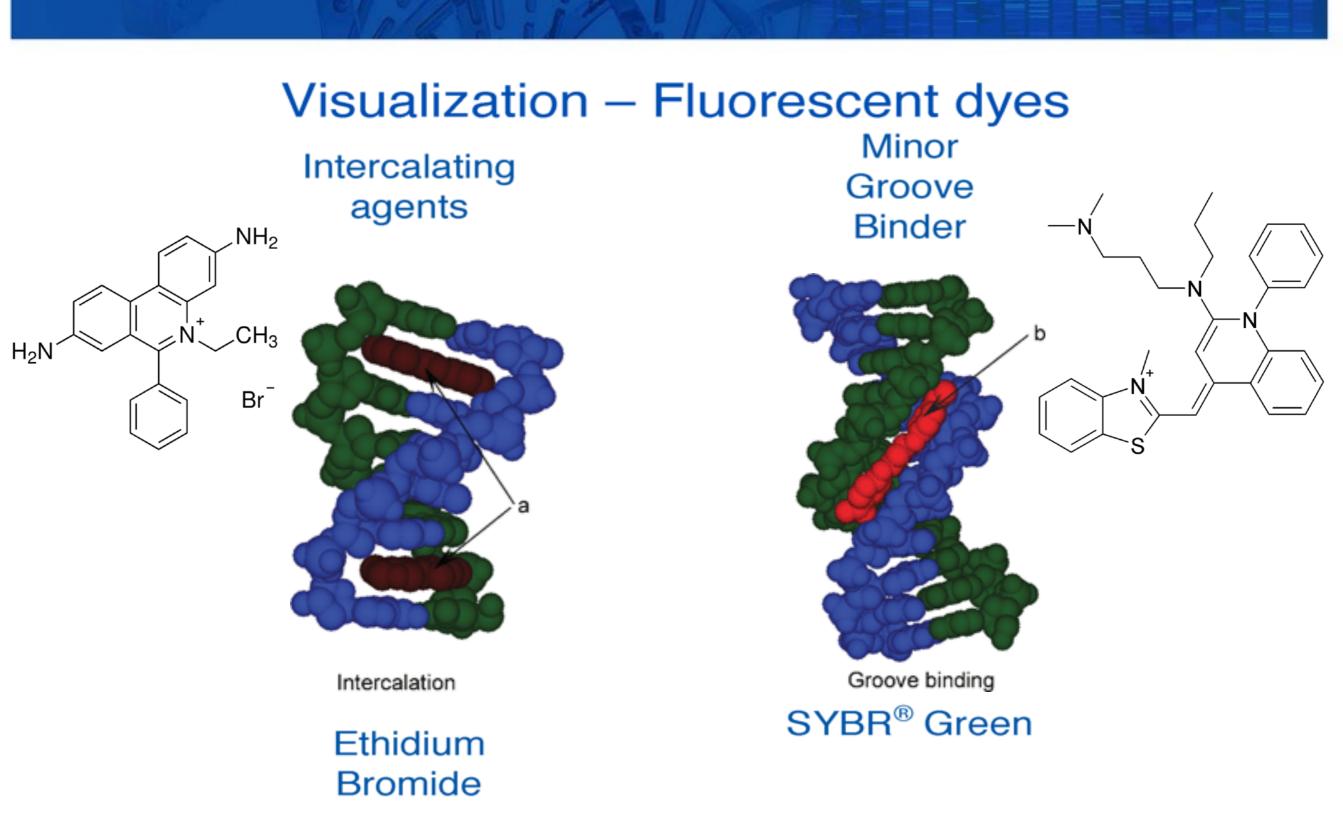
## SYBR<sup>®</sup> Green

# TaqMan<sup>®</sup>

MGB

ROX<sup>™</sup>

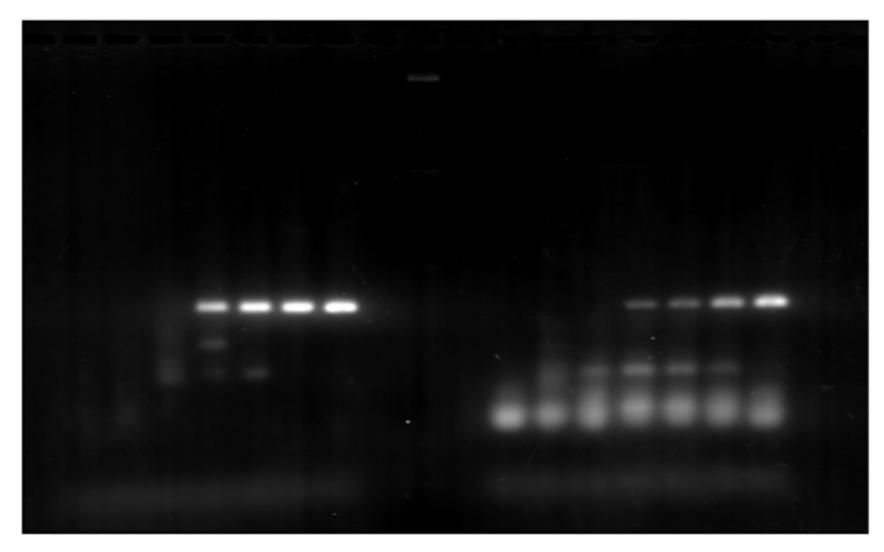
Multicomponenting





### Problem with DNA-binding Dyes

### Bind non-specifically to any double-stranded DNA

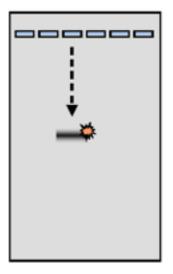


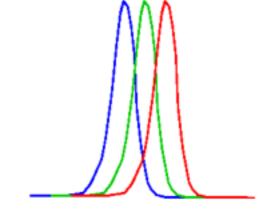
Therefore specificity of the amplifications must be checked



# Exact position of band affected by:

Exact position of peak affected by:



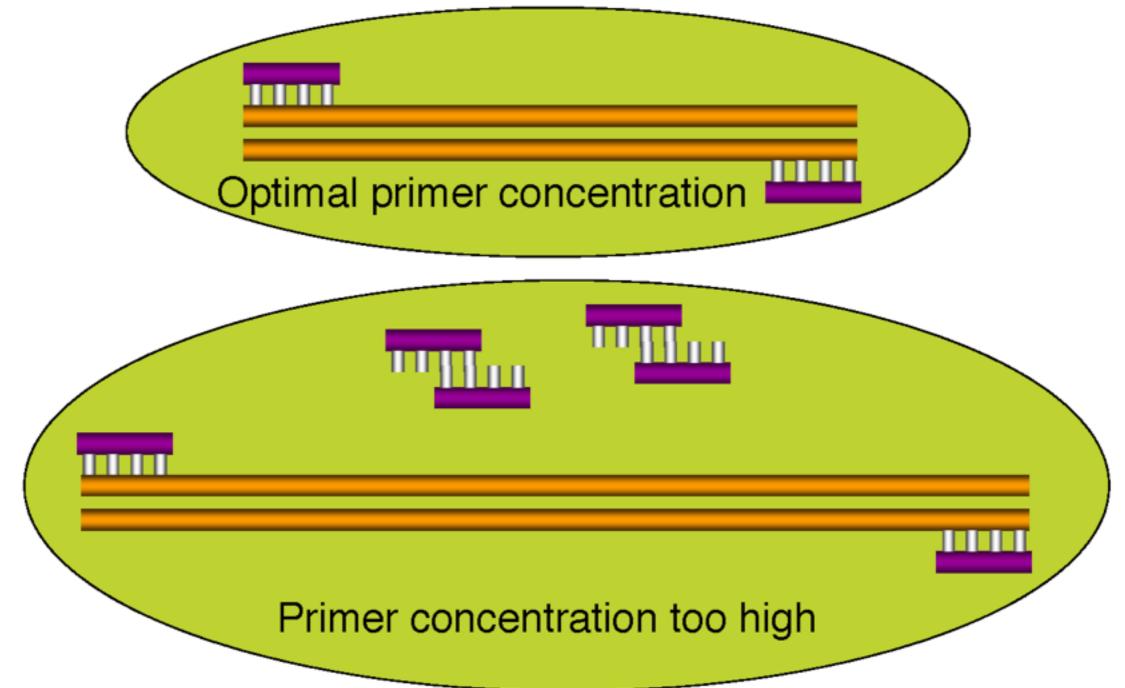


#### 1) size of fragment

1) size of fragment

2) nucleotide composition







#### Primer-dimer formation reduced by minimizing primer concentration

 50 nM
 100 nM
 200 nM
 300 nM
 400 nM
 500 nM

 50 nM
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#### **Forward Primer (final conc)**

Reverse Primer (final conc)



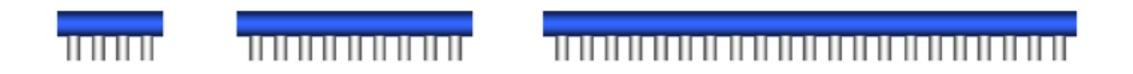
# Real-time PCR SYBR<sup>®</sup> Green TaqMan<sup>®</sup>

### MGB

ROX<sup>™</sup>

## Multicomponenting

# Probing for specific sequence in a pool of very different sequences



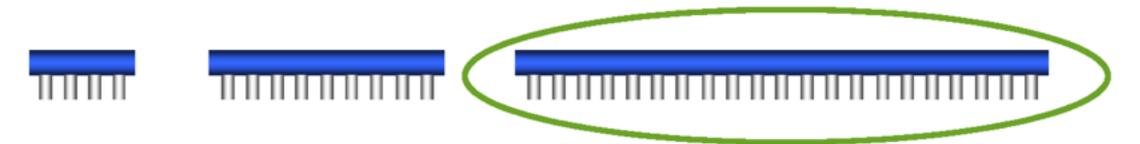
#### 

#### 

#### 

# Probing for specific sequence in a pool of very different sequences

Longer probes increase specificity between different sequences

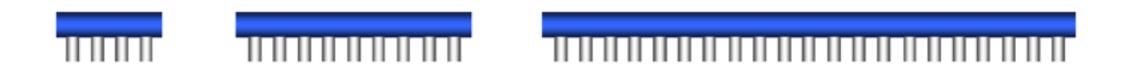


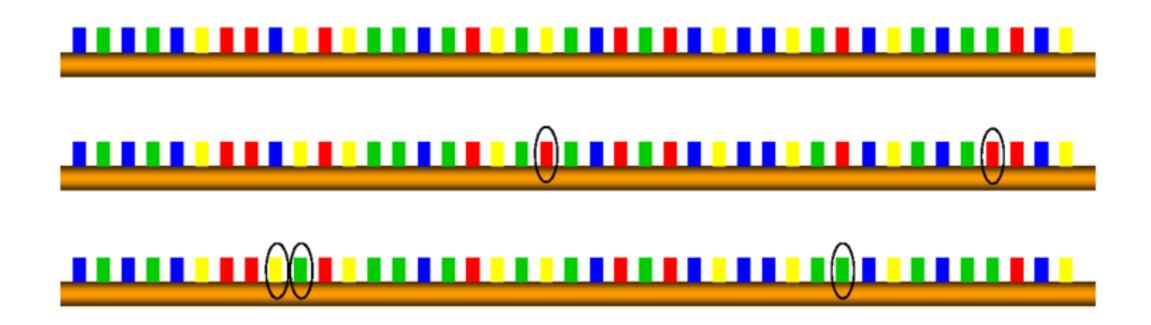
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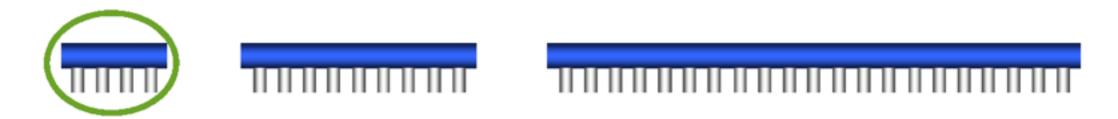
# Probing for specific sequence in a pool of very similar sequences

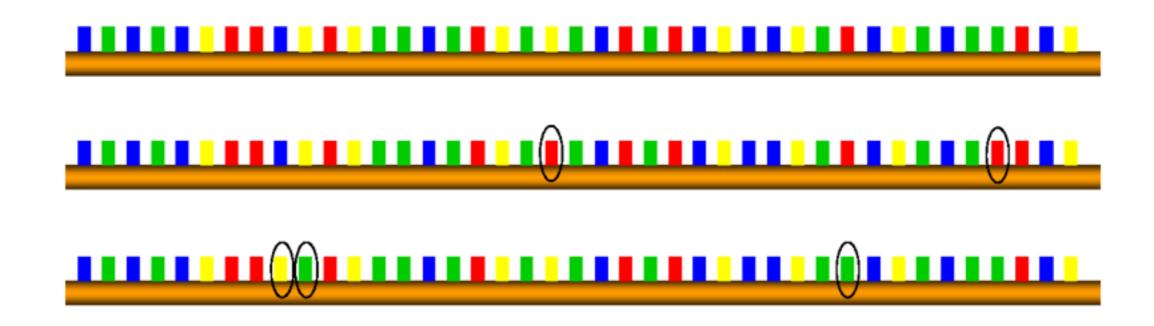




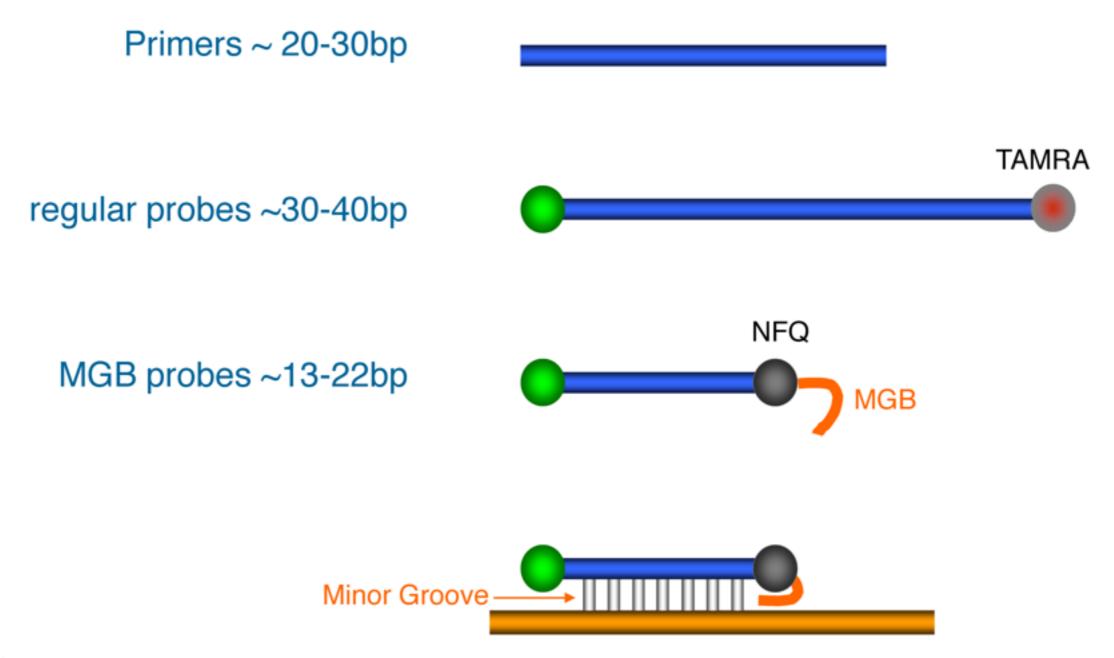
# Probing for specific sequence in a pool of very similar sequences

Shorter probes increase selectivity between similar sequences



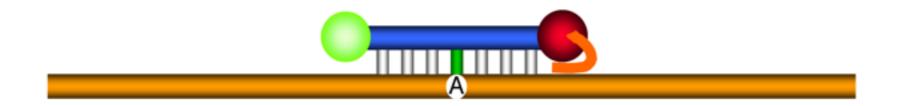


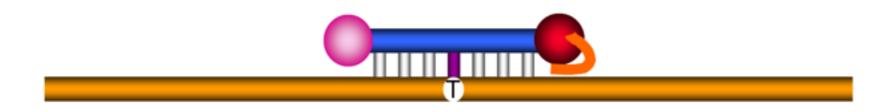
Probe is made shorter by adding a minor-groove-binding molecule that increases probe Tm





### Short MGB probes allow robust single nucleotide specificity ie: SNP assays







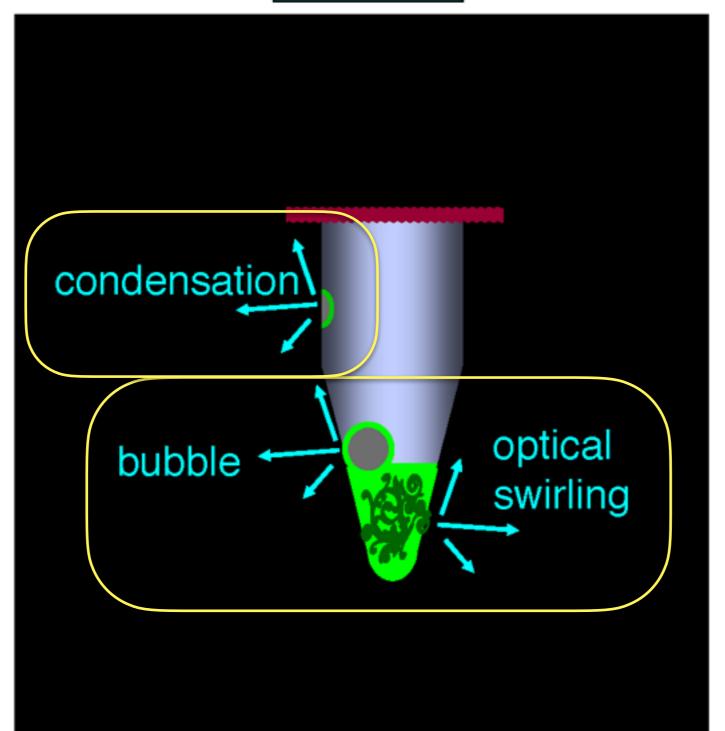
# Real-time PCR SYBR® Green TaqMan<sup>®</sup> MGB

## ROX<sup>™</sup>

## Multicomponenting

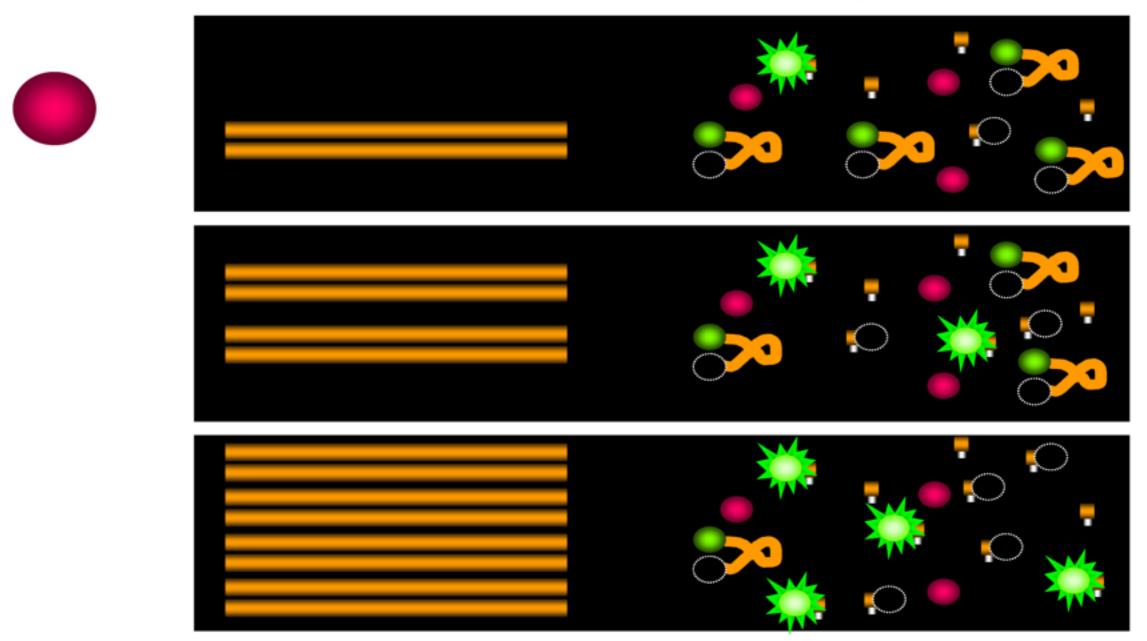


### Common sources of dynamic variation of light signal





### Variation negated by normalizing to a Passive Reference dye

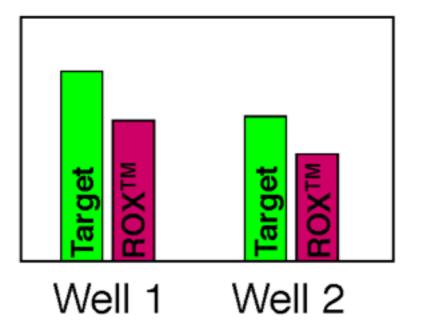


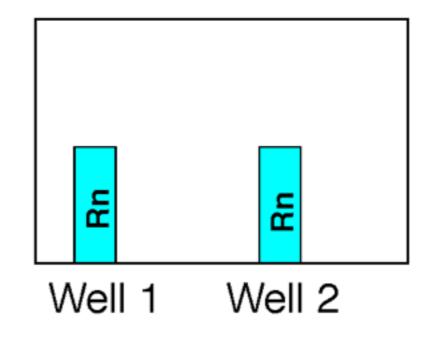


### ROX<sup>™</sup> is a Passive Reference dye

Greatly improves precision of replicates.

Rn = Normalization = Reporter / Reference







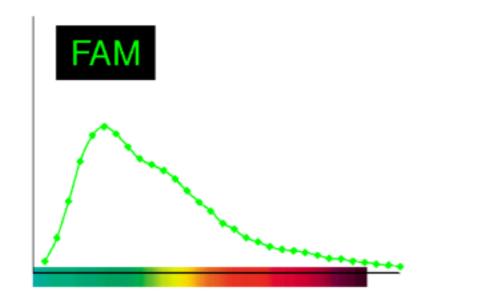
# Real-time PCR SYBR® Green TaqMan<sup>®</sup> MGB

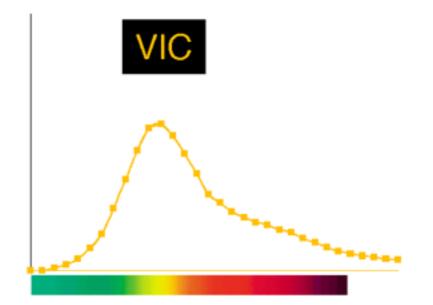
ROX<sup>™</sup>

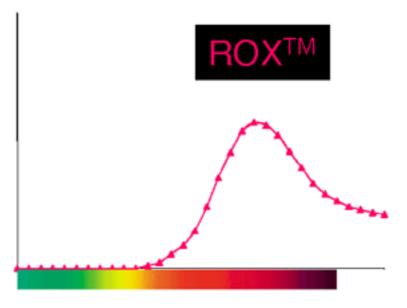
**Multicomponenting** 



Dyes have specific fluorescence spectra with specific peaks

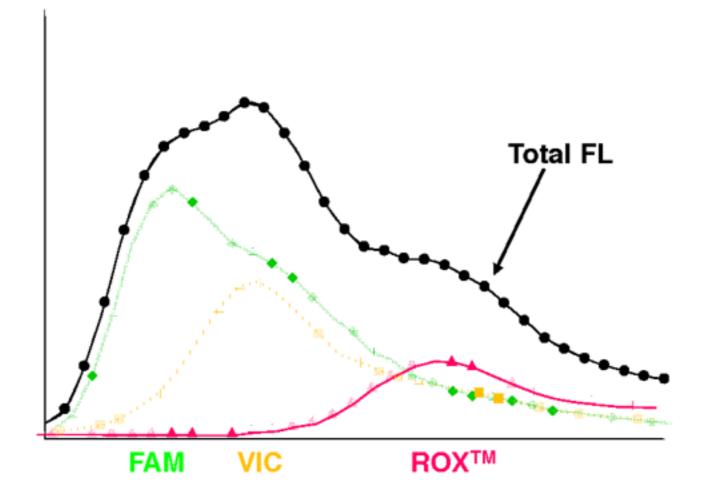








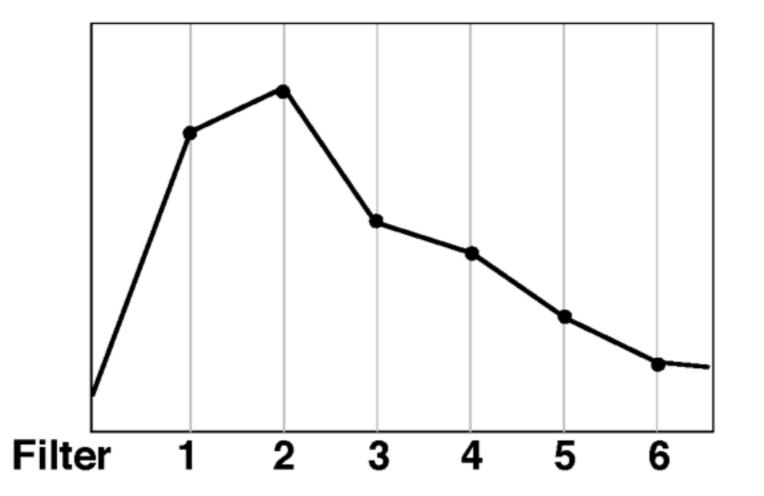
However, if more than one dye is present, there is spectral overlap



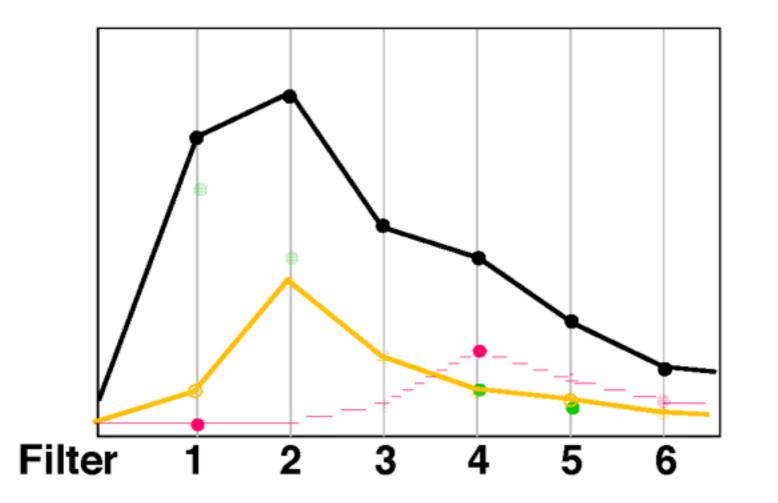
If not addressed, this would introduce large inaccuracies



On the ViiA7, this is depicted as a 5 or 6-point spectral curve

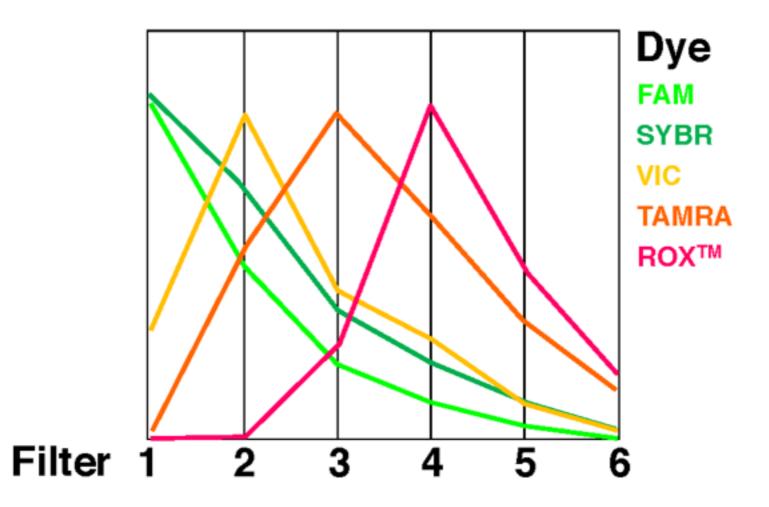


What we see is the Total Fluorescence at each wavelength – this is not the same as the individual dye fluorescence



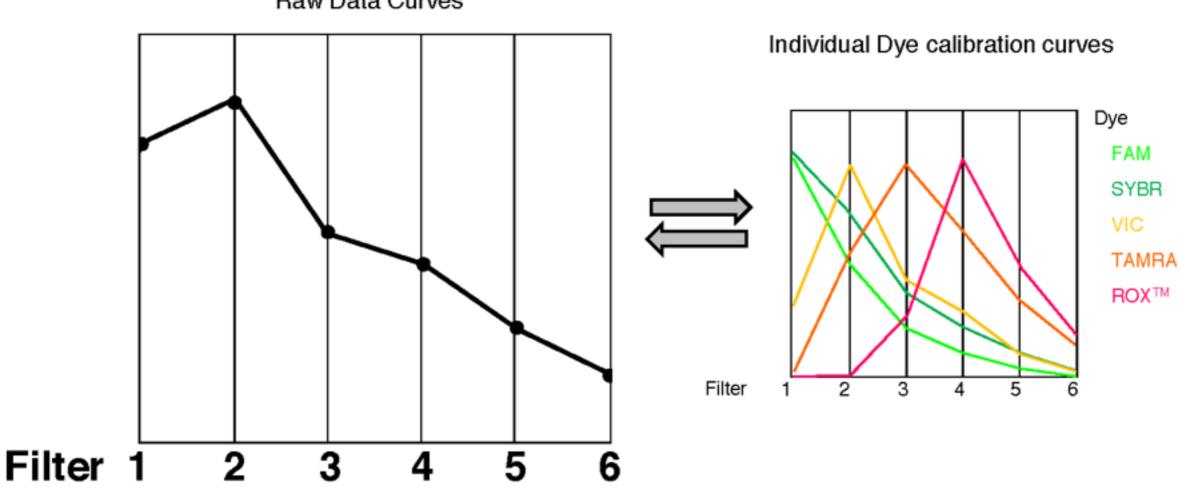
How do we adjust for this?

#### Answer: Dye Calibration

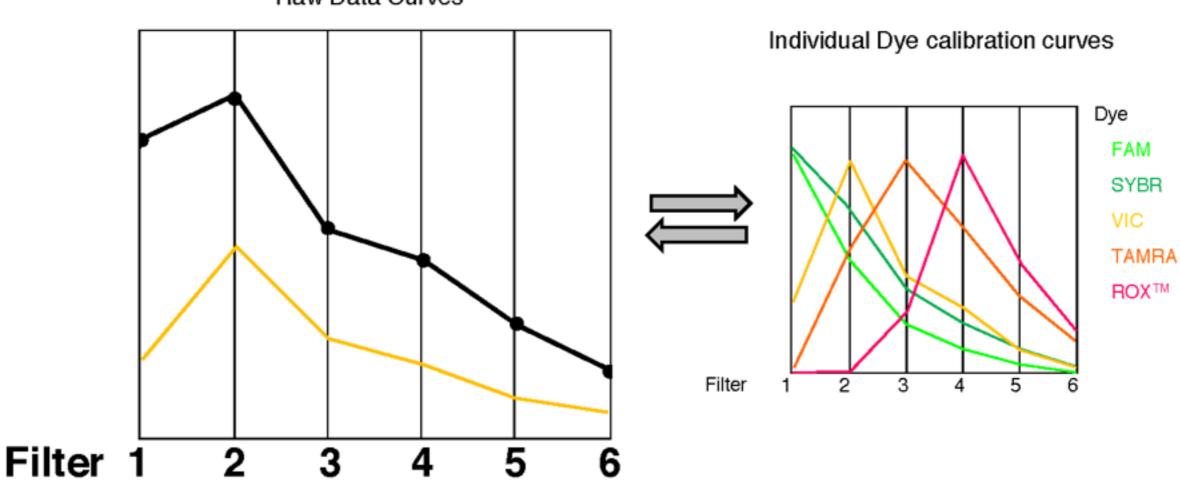


At installation, a dye calibration plate is read. This contains dilutions of pure dye. So the instrument records what each dye "looks like".

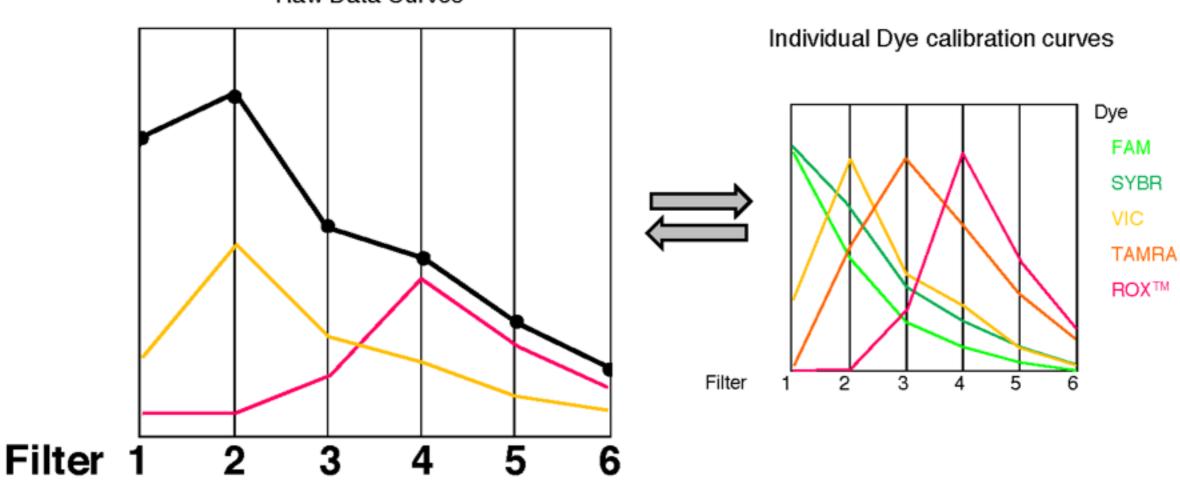






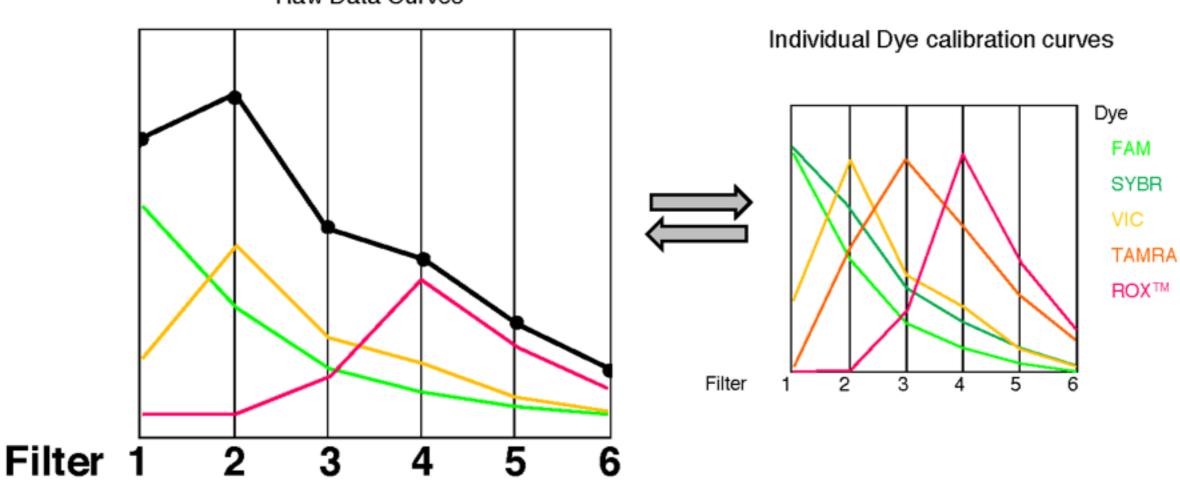


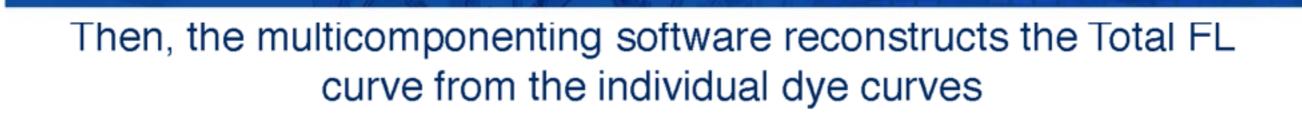


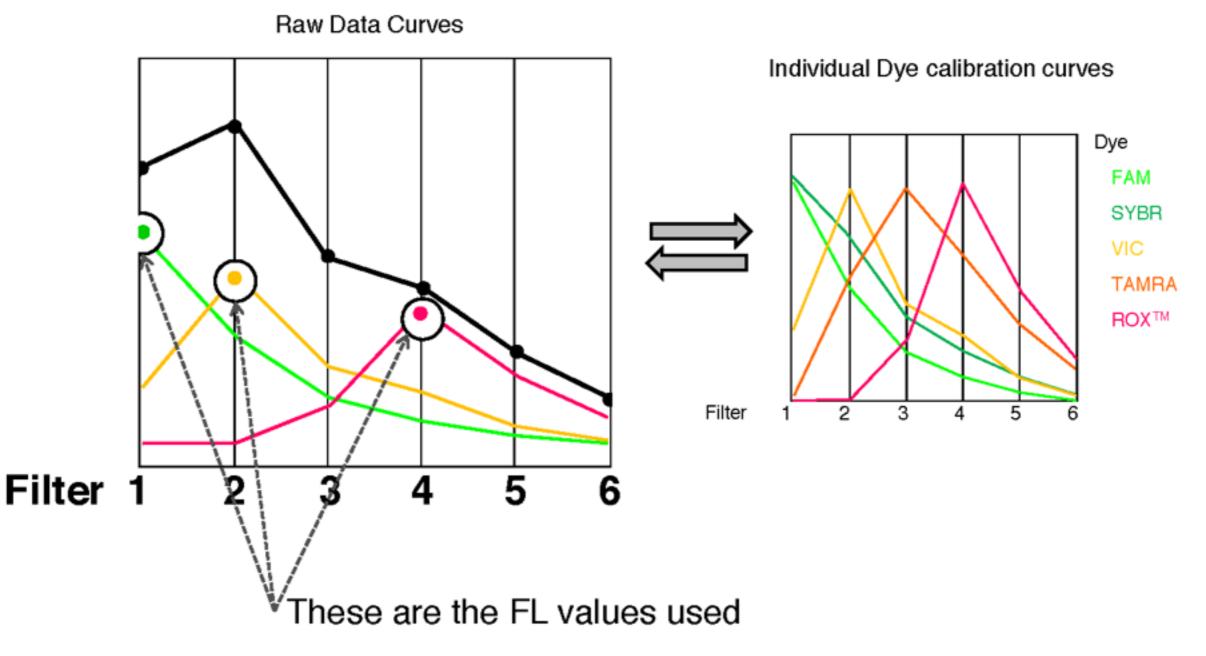


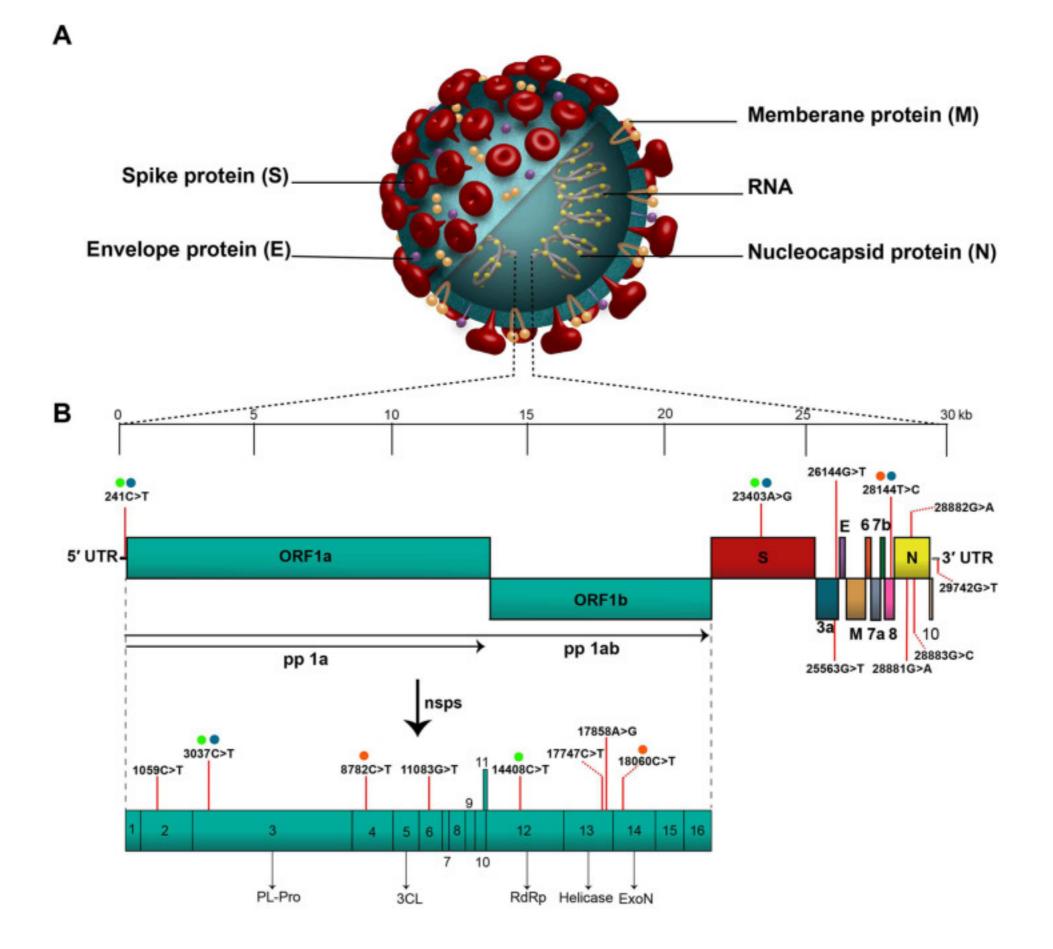


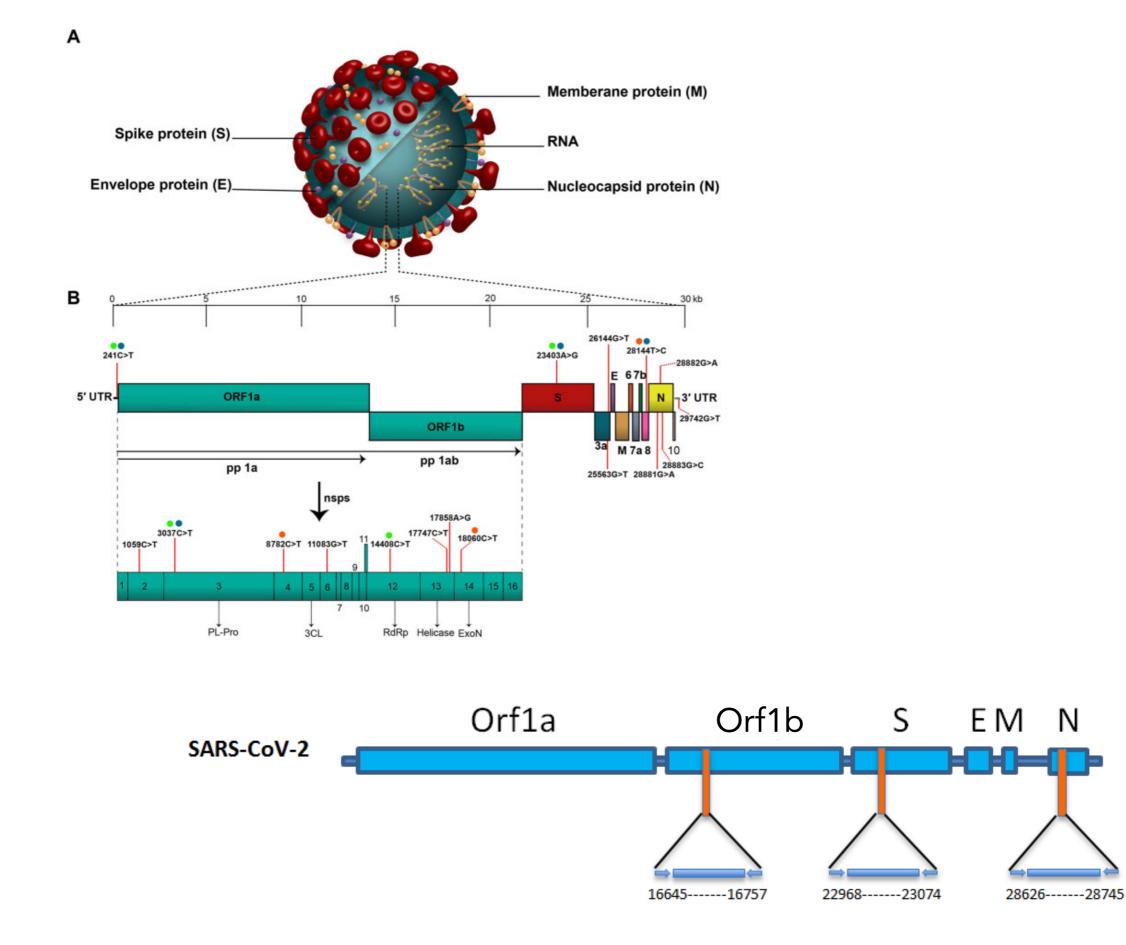
#### Then, the multicomponenting software reconstructs the Total FL curve from the individual dye curves

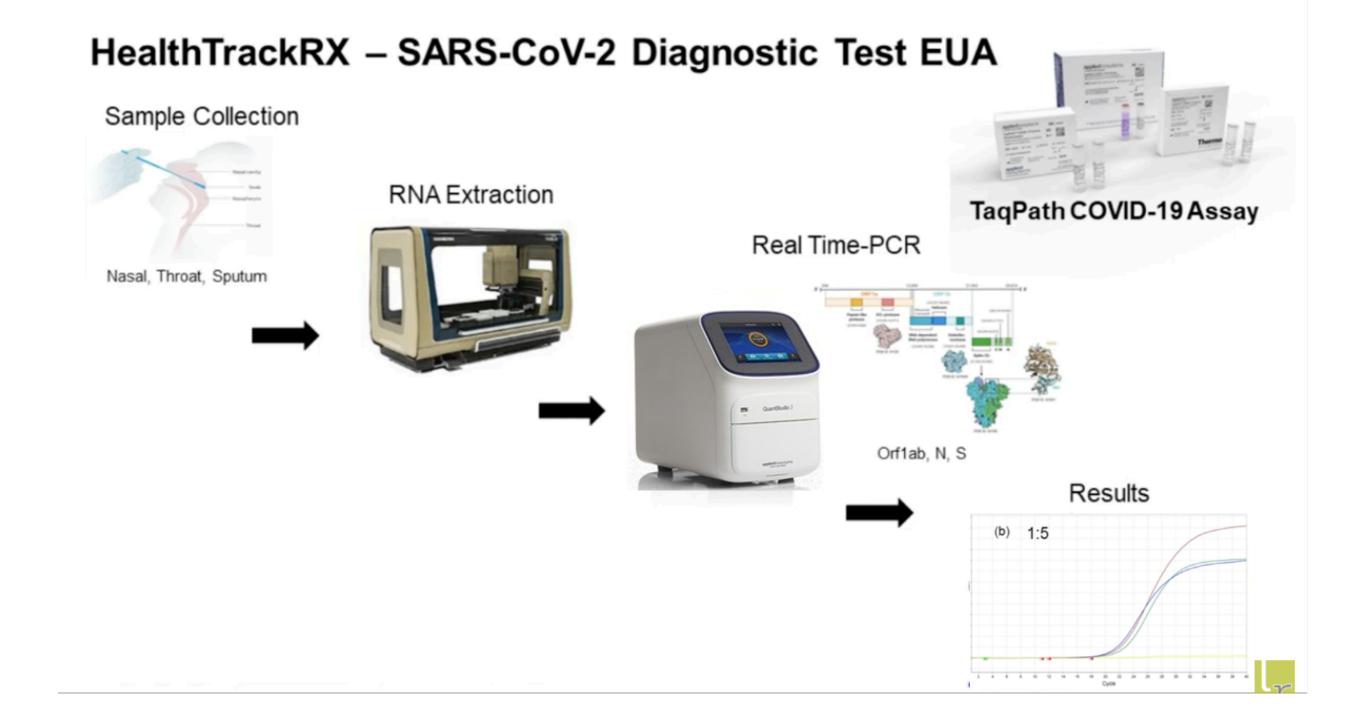


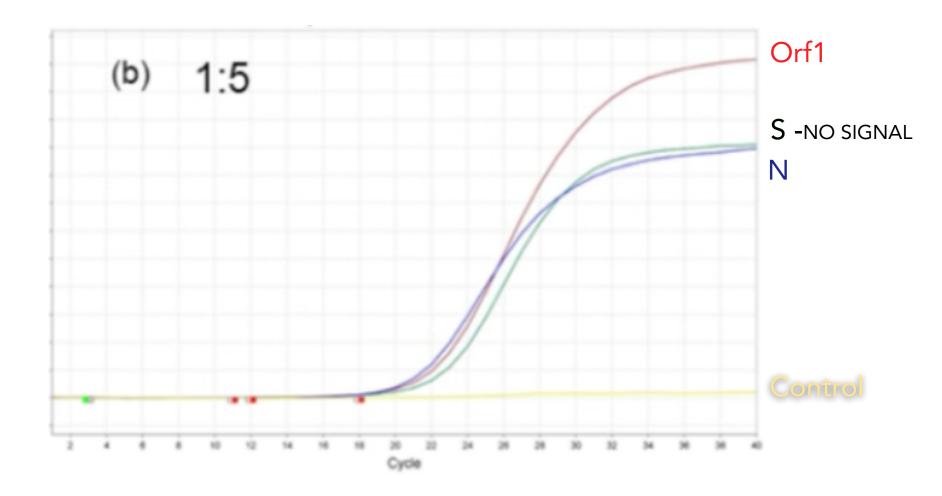


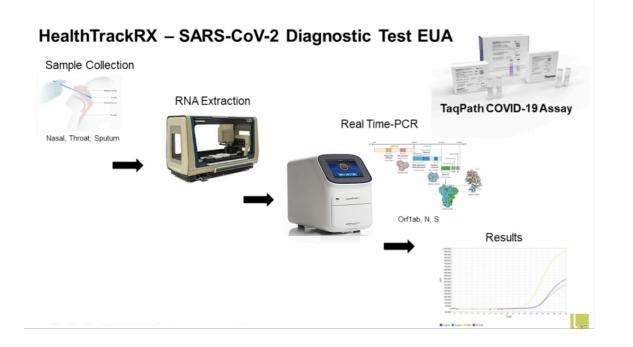












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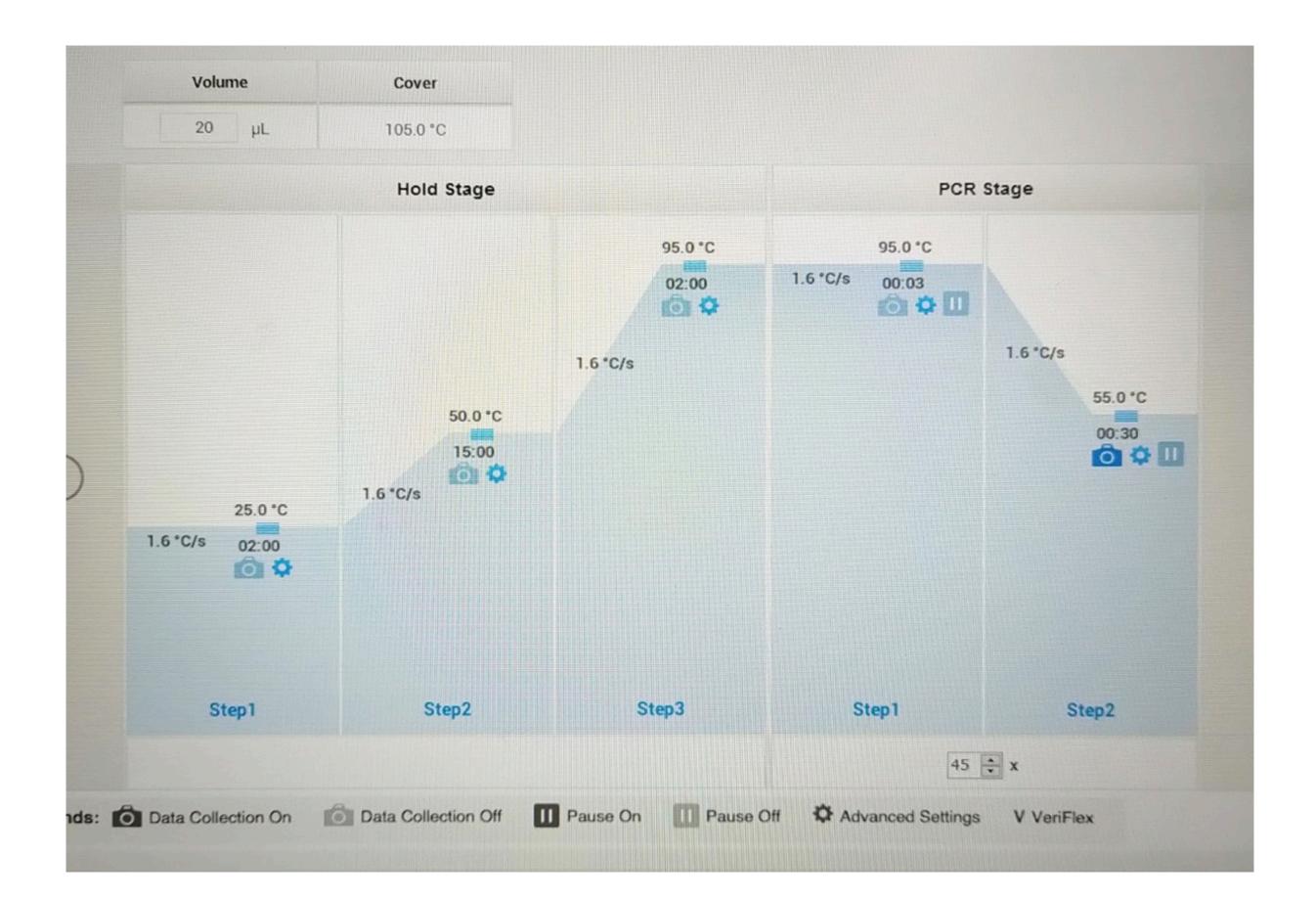
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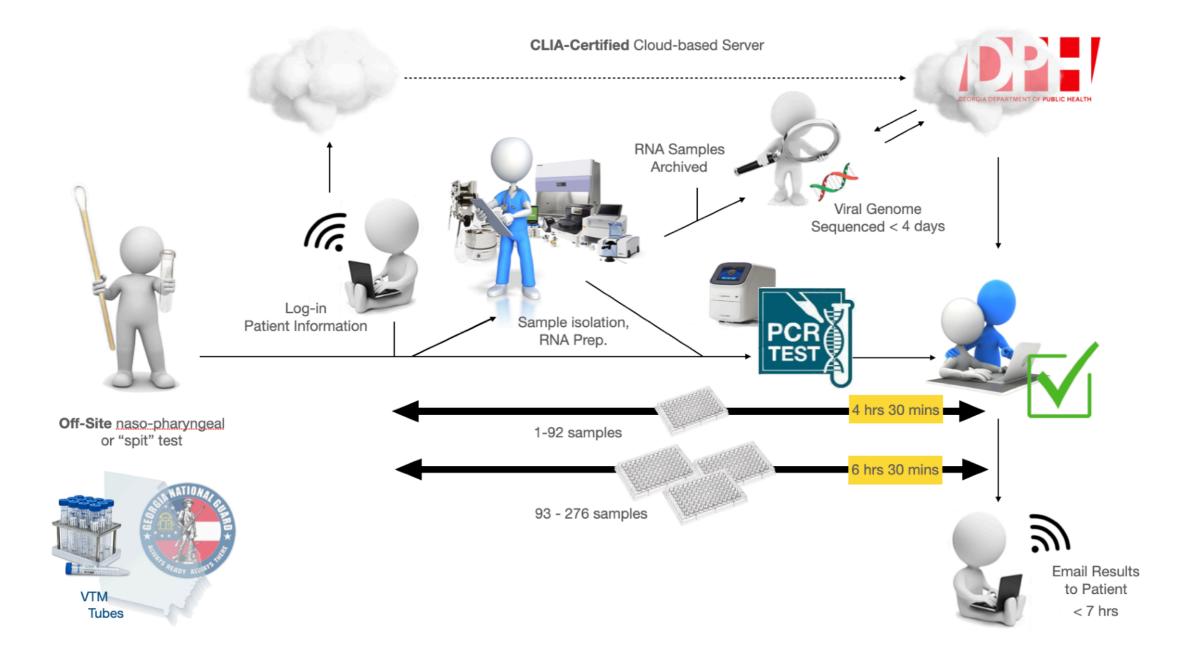
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